Original Article

# The effects of propofol on Doppler echocardiographic values of the aorta, main pulmonary artery, and mitral E wave in New Zealand white rabbits

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#### Abstract

The aim of this study was to determine the effects of propofol on acceleration time (AT), ejection time (ET), AT / ET ratio obtained from the aorta (Ao) and main pulmonary artery (MPA) and Doppler echocardiographic values of mitral E wave (E) in New Zealand white rabbits. Echocardiographic evaluation was performed in 40 conscious rabbits and repeated after propofol administration. The baseline mean values were  $0.028\pm0.006$  second (s) for Ao-AT,  $0.116\pm0.017$  s for Ao-ET,  $0.24\pm0.036$  for Ao-AT / ET,  $0.05\pm0.01$  s for MPA-AT,  $0.137\pm0.018$  s for MPA-ET,  $0.38\pm0.037$  s for MPA-AT/ET,  $84.13\pm18.26$  cm/s for E peak velocity,  $3.06\pm1.38$  mmHg for E peak pressure,  $0.069\pm0.018$  s for E duration,  $0.046\pm0.015$  s for E deceleration time (Edec). After propofol administration, the decrease in velocity (P = 0.044,  $90.76\pm14.39$  vs.  $83.04\pm12.24$ ) and peak pressure gradient (P = 0.036,  $3.47\pm1.01$  vs.  $2.98\pm0.77$ ) of Ao wave, as well as left ventricular ejection fraction (EF) and fractional shortening (FS) was statistically significant (P < 0.001,  $70.1\pm4.47$  vs.  $63.2\pm3.79$ ,  $37.85\pm3.74$  vs.  $32.15\pm2.43$ , respectively); AT was longer (P < 0.05) and AT/ET was increased (P < 0.001) in both Ao and MPA. Among Doppler echocardiographic parameters of the E wave, there was only a statistically significant difference for Edec (P = 0.008). In the presented study, while the effect of propofol is minimal on Edec, its effects on aortic and pulmonic AT/ET values, as well as EF and FS values, should be taken into consideration.

Keywords: acceleration and ejection time, aorta, echocardiography, main pulmonary artery, propofol, rabbit

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#### Introduction

It is observed that cardiovascular diseases are at a significant rate in rabbits, which have become popular pets (Orcutt and Malakoff, 2021). Review articles on cardiac and cardiovascular diseases have been published (Pariaut, 2009; Reusch, 2005; Schnellbacher et al., 2012), and these diseases are discussed in textbooks (Orcutt and Malakoff, 2021; Varga, 2014). The effective use of advanced diagnostic methods such as echocardiography has recently enabled these diseases to be diagnosed more frequently. The prevalence of cardiovascular disease is 2.6%, and the rate of degenerative valve diseases is 40.5% in pet rabbits (Ozawa et al., 2021). In addition, rabbits are generally preferred for experimental degenerative valve disease (Hara et al., 2018; Kim et al., 2023; Lee et al., 2023; Szekeres et al., 2023).

Echocardiographic parameters of mitral flow are commonly used to evaluate the left ventricular (LV) function. One of the echocardiographic parameters used to evaluate diastolic function in human medicine is the measurement of early diastolic mitral inflow wave (E) deceleration time (DT). Shortened or longer Edec is a marker of increased LV chamber stiffness or impaired left ventricular relaxation, respectively (Dori et al., 2014). The ratio of acceleration time to ejection time (AT / ET) is a marker of aortic valve stenosis severity (Einarsen et al., 2021). In the diagnosis of pulmonary hypertension in dogs and cats, the pulmonary flow pattern changes to resemble the aortic flow pattern. This feature is one of the Doppler findings of pulmonary hypertension (Serreset al., 2007).

Propofol is commonly used for the induction and maintenance of anesthesia in both human and veterinary patients. Propofol reduces cardiac output diastolic and systolic blood pressures by causing vasodilation and simultaneously causing a decrease in vascular resistance (Bilotta et al., 2001; Larsen et al., 2007). Propofol has a dose-dependent myocardial depressive effect in vitro studies (Chen et al., 2006; Hamilton et al., 2000) but a minimal effect on echocardiographic Doppler parameters in humans (Yang et al., 2013) and cats (Ferasin, 2009). Although the effects of sedative and anesthetic drugs on cardiac function and echocardiographic parameters have been investigated in pet and laboratory animals (Baumgartner et al., 2010a, b; Marques et al., 2020; Farag et al., 2022; Sandez et al., 2022; Bagardi et al., 2023; Marangoni et al., 2023; Bitencourt et al., 2024; Bockay et al., 2024), the effects of propofol on the Doppler echocardiographic values of the aorta and main pulmonary artery and the deceleration time of the mitral E wave (Edec) have not been reported in rabbits.

This study aimed to determine the effect of propofol on aortic and pulmonic AT, ET, and AT / ET and mitral Edec in New Zealand white rabbits.

# Materials and Methods

The experimental protocol on the use of animals was approved by the Akdeniz University Animal Care Ethics Committee (No: 1653/2023.11.010/112). All animals were normal based on a physical examination and echocardiography. Rabbits were considered normal if they were free of any murmur, extra heart

sounds, arrhythmia, abnormal respiratory sounds on auscultation, and no evidence of any morphologic or hemodynamic cardiac changes on echocardiography such as mitral valve thickening or prolapse, presence of a mitral, tricuspid, aortic or main pulmonary artery regurgitating jet, pericardial or pleural effusion.

Complete echocardiographic examinations, which included transthoracic 2-dimensional (2D), M-mode, and Doppler imaging, were performed by the same observer (MK), more than 20 years' veterinary echocardiography experience, were performed using an ultrasonographic unit (Mindray DC-80, Shenzhen Mindray Bio-medical Electronics, China) equipped with an electronic phased-array sector transducer (8-12 MHz). Echocardiographic parameters were obtained from 40 conscious animals (Group C). Propofol (15 mg/kg, *iv slowly*, Sandoz® İlaç San. Tic. AŞ, İstanbul, Türkiye) was administered to the same animals, and the same echocardiographic parameters were measured within 10 minutes from the beginning of sedation (Group P).

Rabbits were restrained in right and left lateral recumbencies over an opening in an echocardiographic scanning table. An assistant placed the thumb of one hand on the cervicothoracic junction and the other fingers proximal to the elbow joint to direct the forelimbs cranially. The thumb of the other hand was placed just cranial to the iliac crest, and the other fingers proximal to the iliac crest, and the other fingers proximal to the knee joint to direct the hindlimbs caudally. Thus, the rabbits were restrained. Sonographic images of the heart were obtained by placing the ultrasound probe through this opening into the shaved area from the third to fifth intercostal space on the right side and approximately the fourth intercostal space on the left side.

Echocardiographic measurements were obtained according to previously described in rabbits (Giraldo et al., 2019; Turner Giannico et al., 2015). M-mode and spectral Doppler measurements were performed by setting the sweep speed to the fastest level. Since the ECG marker was not used during spectral measurements, mitral A wave and E/A ratio were not evaluated. The following measurements and calculations were carried out: on a right parasternal short-axis view (RPSA) at the level of the papillary muscles, 2D guided M-mode of the interventricular septum, left ventricle (LV) posterior wall, LV internal diameter in diastole and systole, and then the LV ejection fraction (EF) and fractional shortening (FS) (EF% and FS% were calculated from measurements for using the internal ultrasound system software) (Fig. 1); on a RPSA at the level of the aortic valve, peak velocity (Vmax), peak pressure gradient (PGmax), acceleration time (AT), ejection time (ET), AT / ET ratio from MPA flow (Fig. 2); on a left sided parasternal apical (LSPA) five-chamber view, Vmax, PGmax, AT, ET, AT / ET ratio from Ao flow (Fig. 3); LSPA four-chamber view, peak velocity of E wave (Vvel), Edec, E duration (Edur) (Fig. 4). The measurement of HR was carried out manually using the echocardiographic software for each loop individually (Fig. 1).

Statistical analysis was carried out on commercial software (IBM SPSS Statistics 22.0, SPSS Inc., USA). Data were expressed as the mean ± standard deviation (SD). A two-sample *t*-test for all echocardiographic

parameters was used to assess the differences between groups. A P value < .05 was regarded as statistically significant.

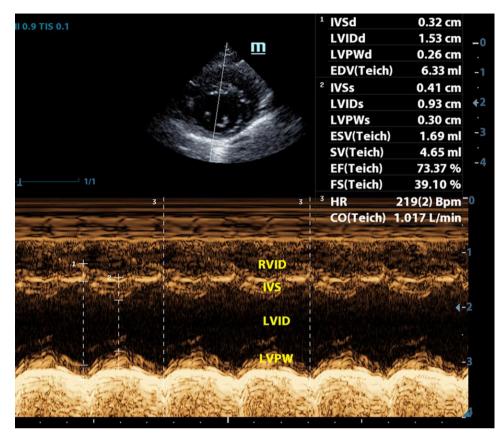


Figure 1 From the right parasternal short axis at the level of papillary muscles, measurement of echocardiographic M-Mode parameters of the left ventricle. RVID: right ventricle internal diameter, IVS: interventricular septum, LVID: left ventricle internal diameter, LVPW: left ventricle posterior wall, EF: ejection fraction, FS: fractional shortening, HR: heart rate, CO: cardiac output.

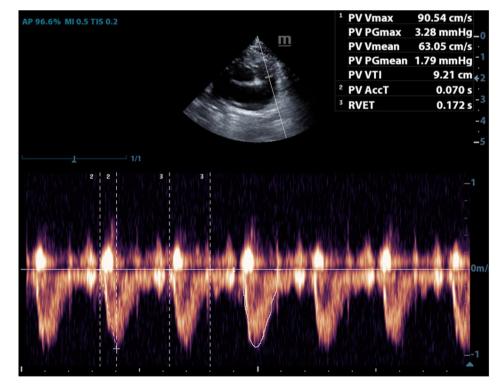


Figure 2 From the right parasternal short axis at the level of the aorta, measurement of echocardiographic Doppler parameters of the main pulmonary artery. Vmax: peak velocity; PGmax: peak pressure gradient; PV AccT: Acceleration time (AT); RVET: right ventricle ejection time (ET). The AT / ET ratio is 0.407 in this rabbit.

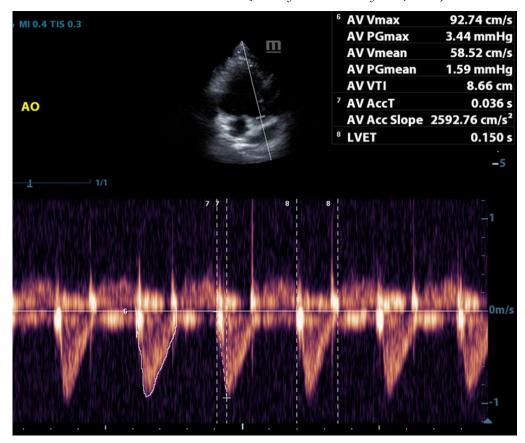
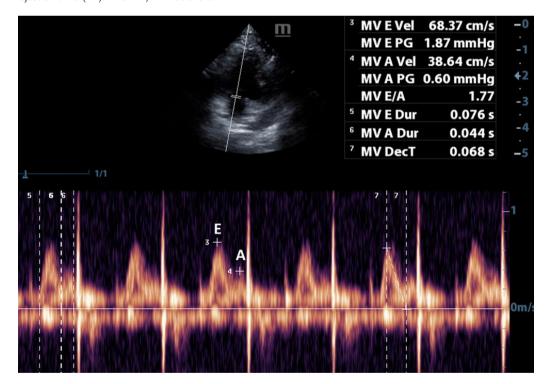


Figure 3 From the left side of the parasternal apical five-chamber view, the echocardiographic Doppler parameters of the aorta are measured. Vmax: peak velocity; PGmax: peak pressure gradient; AV AccT: Acceleration time (AT); LVET: left ventricle ejection time (ET). The AT / ET ratio is 0.24.



**Figure 4** From the left side of the parasternal apical five-chamber view, the echocardiographic Doppler parameters of the mitral flow in a rabbit are measured. E vel: peak velocity of the E wave, E PG: peak pressure gradient of the E wave, E Dur: duration of the E wave, DecT: deceleration time of E wave.

#### Result

In total, the echocardiographic parameters of 40 conscious and sedated healthy male New Zealand white rabbits were analyzed. Mean age and mean BW were  $12.65\pm1.89$  months (10-16 months) and  $3.38\pm0.36$  kg (2.7-3.8 kg), respectively. No complications associated with propofol were encountered in any animals, and no mortality occurred.

Table 1 shows the echocardiographic values measured from flows of the aorta and main pulmonary artery in Group C. The HR, EF%, and FS% values were lower in Group P than in Group C (P < 0.001, 197.55  $\pm$  18.8 bpm vs 239.43  $\pm$  26.62 bpm, 63.2  $\pm$  3.79 vs 70.1  $\pm$  4.47, 32.15  $\pm$  2.43 vs 37.85  $\pm$  3.74, respectively).

Peak velocities and pressures of Ao were decreased in Group P (P < 0.05). While the pulmonic flow pattern was substantially symmetrical (Fig. 2), the aortic flow pattern was asymmetric in Group C (Fig. 3). In both aortic and pulmonic flow patterns, the AT was longer (P < 0.05), and thus pulmonic flow pattern became more symmetrical in Group P. The change in the aortic and pulmonic AT/ET was statistically significant (P < 0.001) (Table 2).

Administration of propofol did not change the mitral flow pattern. Among Doppler echocardiographic values of the mitral E wave, there was only a statistically significant difference for Edec (P = 0.008) (Table 3).

**Table 1** Mean, 95% confidence interval (CI), and minimum-maximum values of echocardiographic values from flows of the aorta and main pulmonary artery in 40 conscious New Zealand white rabbits.

Vessel	Parameters	Mean ± SD	95% CI	Range
	Vmax (cm/s)	$90.76 \pm 14.39$	86.16-95.36	63.5-127.4
	PGmax (mmHg)	$3.47 \pm 1.01$	3.15-3.79	1.61-6.5
Ao	AT(s)	$0.028 \pm 0.006$	0.026-0.03	0.018-0.035
	ET (s)	$0.116 \pm 0.017$	0.112-0.123	0.096-0.154
	AT/ET	$0.23 \pm 0.036$	0.22-0.25	0.18-0.30
	Vmax (cm/s)	$83.42 \pm 13.29$	78.85-87,99	61.3-112.4
	PGmax (mmHg)	$2.85 \pm 0.97$	2.54-3.17	1.5-5.05
MPA	AT(s)	$0.05 \pm 0.01$	0.049-0.055	0.03-0.07
	ET (s)	$0.137 \pm 0.018$	0.131-0.143	0.099-0.174
	AT/ET	$0.38 \pm 0.037$	0.37-0.4	0.33-0.49

Abbreviations: SD, standard deviation; Ao, Aorta; MPA, Main pulmonary artery; Vmax, peak velocity; PGmax, peak pressure gradient; AT, acceleration time; ET, ejection time; AT/ET: ratio of AT to ET.

**Table 2** Comparison of Doppler echocardiographic parameters from flows of the aorta and main pulmonary artery between groups.

Vessel	Parameters	Group	Mean ± SD	P value
	Vmax (cm/s)	С	90.76 ± 14.39	0.044
	vinax (cm/s)	P	$83.04 \pm 12.24$	0.044
	DC may (www.Ha)	C	$3.47 \pm 1.01$	0.036
	PGmax (mmHg)	P	$2.98 \pm 0.77$	0.036
Ao	<b>A</b> T (-)	C	$0.028 \pm 0.006$	0.04
A0	AT(s)	P	$0.032 \pm 0.005$	0.04
	ET (a)	C	$0.116 \pm 0.017$	0.208
	ET (s)	P	$0.111 \pm 0.015$	0.208
	AT / ET	С	$0.23 \pm 0.036$	0.001
		P	$0.28 \pm 0.021$	
	Venan (aur/a)	С	83.42 ± 13.29	0.245
	Vmax (cm/s)	P	79.74 ± 12.74	0.345
	PGmax (mmHg)	С	$2.85 \pm 0.97$	0.322
	1 Gillax (mm11g)	P	$2.6 \pm 0.85$	0.322
MPA	AT (s)	С	$0.05 \pm 0.001$	0.037
IVII A	AT (S)	P	$0.056 \pm 0.007$	0.037
	ET (s)	C	$0.137 \pm 0.018$	0.639
		P	$0.135 \pm 0.016$	
	AT / ET	С	$0.38 \pm 0.037$	0.001
		P	$0.44 \pm 0.036$	0.001

Abbreviations: SD, standard deviation; CI, confidence interval; C, conscious; P, propofol; Ao, aorta; MPA, main pulmonary artery; Vmax, peak velocity; PGmax, peak pressure gradient; AT, acceleration time; ET, ejection time; AT/ET, the ratio of AT to ET.

**Table 3** Comparison of Doppler echocardiographic parameters of the mitral E wave between groups.

Parameters	Group	Mean ± SD	P value
Evel (cm/s)	С	$84.13 \pm 18.26$	0.054
Ever (cm/s)	P	77.6 ± 15.99	
EDC (mmHa)	С	$3.06 \pm 1.38$	0.064
E PG (mmHg)	P	$2.69 \pm 1.32$	
E1 ()	С	$0.069 \pm 0.018$	0.059
Edur (s)	P	$0.06 \pm 0.014$	
E1 ()	С	$0.046 \pm 0.015$	0.000
Edec (s)	P	$0.034 \pm 0.009$	0.008

### Discussion

Based on a review of the literature, this is the first study proposing the reference intervals for AT, ET, and AT / ET from flows of the aorta and main pulmonary artery in NZW rabbits. Aortic AT / ET is a straightforward and reproducible echocardiographic parameter that combines the severity of aortic stenosis with the effects it has on the left ventricle. AT / ET > 0.35 is a strong predictor of outcome in human patients with high-gradient severe aortic stenosis (Altes et al., 2021). Pulmonic AT, ET, and AT / ET are used to evaluate both humans and dogs with precapillary pulmonary hypertension (PH). Three different patterns of flow have been described: type I: normal, symmetrical flow pattern; type II: moderate PH, asymmetrical flow pattern; shortened AT; type III, severe PH, furthermore shortened AT and mid-systolic notch in the flow pattern (Atkinson et al., 2009; Serres et al., 2007). The threshold value for predicting PH in dogs has been reported as pulmonic AT / ET < 0.31 and AT < 0.058 seconds (s) (Schober and Baade, 2006). In a cat with type III reversible pulmonary hypertension, pulmonic AT, ET, and AT / ET have been reported as 0.04 s, 0.133 s, and 0.26, respectively (Baron Toaldo et al., 2011). In normal dogs, the aortic flow pattern is typically asymmetric, and AT / ET is lower than 0.3, whereas the pulmonic flow pattern has a more symmetric shape and an AT/ET greater than 0.43 (Kirberger et al., 1992). Although the aortic AT / ET in the presented study was consistent with the values obtained in dogs, pulmonic AT / ET was lower than the values in dogs. On the other hand, pulmonic AT / ET (median: 0.37, range: 0.28-0.41) in healthy cats (Lachance et al., 2022) is consistent with the findings of this study.

The use of a sedative protocol to reduce stress and facilitate handling in rabbit echocardiography is common, especially in experimental studies. The combination frequently used for this purpose is ketamine- $\alpha_2$  agonists. This combination has the potential for cardiac and respiratory depression. The combinations of ketamine-midazolam (Fontes-Sousa et al., 2009) and dexmedetomidine-midazolam-morphine (Bitencourt et al., 2023) have been reported to be associated with minimal cardiorespiratory depression. In low-risk human patients, propofol causes a negligible subclinical reduction in LV systolic and diastolic echocardiographic parameters (Yang et al., 2013). Although this anesthetic caused a slight decrease in myocardial systolic velocities, a tissue Doppler study showed that it did not affect diastolic function in cats (Ferasin, 2009). In the presented study, although propofol was effective on LV systolic function parameters (EF% and FS%) and HR, it did not cause systolic dysfunction or bradycardia, and thus its effect appears to be minimal on LV systolic function, as in humans (Freitas et al., 2022). In the presented study, the decrease in FS and EF values after propofol administration can be explained by a decrease in cardiac output (Sahinovic et al., 2018). Additionally, Doppler echocardiographic values of mitral E wave were not affected, except for Edec. The decrease in both aortic and pulmonary velocities and pressure gradients seen after propofol administration in this study may be due to partial suppression of left ventricular (LV) systolic function (Freitas et~al., 2022) and a significant decrease in sympathetic tone. (Sahinovic et~al., 2018). Although the increase in the pulmonic AT / ET makes this flow pattern more symmetrical, it is a situation that should be taken into account in future studies. The mean aortic AT / ET (0.29  $\pm$  0.021) in Group P is a change that should be taken into account, although the fact that it is below the threshold value for human aortic stenosis > 0.35 may also indicate a difference that does not cause adverse effects. Further research is now needed to determine the Doppler echocardiographic diagnostic criteria of aortic stenosis and pulmonary hypertension in rabbits.

As a result, propofol administration may not have any effect on Doppler echocardiographic values of mitral E wave, except for Edec. The effect of propofol on aortic and pulmonic AT / ET, as well as LV systolic function parameters, should be taken into account.

## Acknowledgment

The experimental protocol on the use of animals was approved by the Akdeniz University Animal Care Ethics Committee (No: 1653/2023.11.010/112). The authors confirm that they have adhered to ARRIVE Guidelines to protect animals used for scientific purposes.

**Conflict of interest:** The authors declare no conflict of interest.

# References

Altes A, Thellier N, Bohbot Y, Ringle Griguer A, Verdun S, Levy F, Castel AL, Delelis F, Mailliet A, Tribouilloy C and Maréchaux S 2021. Relationship Between the Ratio of Acceleration Time/Ejection Time and Mortality in Patients With High-Gradient Severe Aortic Stenosis. J Am Heart Assoc. 10(23): e021873. doi: 10.1161/JAHA.121.021873.

Atkinson KJ, Fine DM, Thombs LA, Gorelick JJ and Durham HE 2009. Evaluation of pimobendan and N-terminal pro-brain natriuretic peptide in the treatment of pulmonary hypertension secondary to degenerative mitral valve disease in dogs. J Vet Intern Med. 23(6): 1190-1196. doi: 10.1111/j.1939-1676.2009.0390.x.

Bagardi M, Locatelli C, Brambilla PG, Ghilardi S, Rabbogliatti V, Amari M, Casiraghi S, Ravasio G, Galimberti L and Brioschi FA 2023. Comparison of two multimodal intramuscular anaesthetic protocols in uncooperative feline patients: effects on sedation and echocardiographic measurements. J Feline Med Surg. 25(2): 1098612X231153069. doi: 10.1177/1098612X231153069.

Bočkay A, Agudelo CF, Figurová M, Vargová N and Trbolová A 2024. Effect of Butorphanol-Medetomidine and Butorphanol-Dexmedetomidine on Echocardiographic Parameters during Propofol Anaesthesia in Dogs. Animals (Basel). 14(9): 1379. doi: 10.3390/ani14091379.

Baron Toaldo M, Guglielmini C, Diana A, Giunti M, Dondi F and Cipone M 2011. Reversible pulmonary

- hypertension in a cat. J Small Anim Pract. 52(5): 271-277. doi: 10.1111/j.1748-5827.2011.01064.x.
- Baumgartner C, Bollerhey M, Ebner J, Laacke-Singer L, Schuster T and Erhardt W 2010. Effects of ketamine-xylazine intravenous bolus injection on cardiovascular function in rabbits. Can J Vet Res. 74(3): 200-208.
- Baumgartner C, Bollerhey M, Ebner J, Schuster T, Henke J and Erhardt W 2010. Effects of medetomidine-midazolam-fentanyl IV bolus injections and its reversal by specific antagonists on cardiovascular function in rabbits. Can J Vet Res. 74(4): 286-298.
- Bilotta F, Fiorani L, La Rosa I, Spinelli F and Rosa G 2001. Cardiovascular effects of intravenous propofol administered at two infusion rates: a transthoracic echocardiographic study. Anaesthesia. 56(3): 266-271. doi: 10.1046/j.1365-2044.2001.01717-5.x.
- Bitencourt EH, de Lima MPA, Barreto MSO, de Sousa FG, Silva EF, Pimenta ELM and Beier SL 2024. Clinical and Doppler echocardiographic evaluation of rabbits sedated with dexmedetomidine in combination with midazolam and morphine. Ciência Rural. 54: e20220259. doi: 10.1590/0103-8478cr20220259.
- Chen WH, Lee CY, Hung KC, Yeh FC, Tseng CC and Shiau JM 2006. The direct cardiac effect of propofol on intact isolated rabbit heart. Acta Anaesthesiol Taiwan. 44(1): 19-23.
- Dori G, Egbaria MA and Jabaren M 2014. Two-slope ascending arm of the early trans-mitral flow velocity Doppler wave in patients with heart failure and preserved ejection fraction. Med Hypotheses. 83(6): 735-739. doi: 10.1016/j.mehy.2014.09.030.
- Einarsen E, Cramariuc D, Bahlmann E, Midtbo H, Chambers JB and Gerdts E 2021. Higher Acceleration/Ejection Time Ratio Predicts Impaired Outcome in Aortic Valve Stenosis. Circ Cardiovasc Imaging. 14(1): e011467. doi: 10.1161/CIRCIMAGING.120.011467.
- Farag A, Mandour AS, Hamabe L, Yoshida T, Shimada K and Tanaka R 2022. Novel protocol to establish the myocardial infarction model in rats using a combination of medetomidine-midazolambutorphanol (MMB) and atipamezole. Front Vet Sci. 5(9): 1064836. doi: 10.3389/fvets.2022.1064836.
- Ferasin L 2009. Feline myocardial disease 2: diagnosis, prognosis and clinical management. J Feline Med Surg. 11(3): 183-194. doi: 10.1016/j.jfms.2009.01.002.
- Fontes-Sousa AP, Moura C, Carneiro CS, Teixeira-Pinto A, Areias JC and Leite-Moreira AF 2009. Echocardiographic evaluation including tissue Doppler imaging in New Zealand white rabbits sedated with ketamine and midazolam. Vet J. 181(3): 326-331. doi: 10.1016/j.tvjl.2008.02.022.
- Freitas TM, David C, Almeida AG, Pinto FJ, Costa J and Caldeira D 2022. Cardiovascular and Respiratory Safety of Sedation Strategies Used in Transesophageal Echocardiography: A Systematic Review Incorporating Network Meta-Analysis. J Cardiothorac Vasc Anesth. 36(11): 4129-4140. doi: 10.1053/j.jvca.2022.07.003.
- Giraldo A, Talavera López J, Brooks G and Fernández-Del-Palacio MJ 2019. Transthoracic

- echocardiographic examination in the rabbit model. J Vis Exp. 1(148): e59457. doi: 10.3791/59457.
- Hamilton DL, Boyett MR, Harrison SM, Davies LA and Hopkins PM 2000. The concentration-dependent effects of propofol on rat ventricular myocytes. Anesth Analg. 91(2): 276-282. doi: 10.1097/00000539-200008000-00005.
- Hara T, Tsukada N, Okano M, Ishida T, Hirata KI and Shiomi M 2018. Progression of calcific aortic valve sclerosis in WHHLMI rabbits. Atherosclerosis. 273: 8-14. doi: 10.1016/j.atherosclerosis.2018.03.044.
- Kim E, Park EH, Kim JM, Lee E, Park SH, Kim CW, Choi IJ, Oak MH and Chang K 2023. A Rabbit Aortic Valve Stenosis Model Induced by Direct Balloon Injury. J Vis Exp. 31(193): doi: 10.3791/65078.
- Kirberger RM, Bland-van der Berg P and Grimbeek RJ 1992. Doppler echocardiography in the normal dog. Part II. Factors influencing blood flow velocities and a comparison between left and right heart blood flow. Vet Radiol Ultrasound. 33(6): 380-386. doi: 10.1111/j.1740-8261.1992.tb00163.x.
- Lachance L, Conversy B, Wiggen K, Pavard C, Reinero C and Masseau I 2022. No evidence of pulmonary hypertension revealed in an echographic evaluation of right-sided hemodynamics in hyperthyroid cats. J Feline Med Surg. 24(12):e558-e567. doi: 10.1177/1098612X221127102.
- Larsen JR, Torp P, Norrild K and Sloth E 2007. Propofol reduces tissue-Doppler markers of left ventricle function: a transthoracic echocardiographic study. Br J Anaesth. 98(2): 183-188. doi: 10.1093/bja/ael345.
- Lee GH, Kang KK, Yun HH, Jo W and Oh T 2023. Multiple time-dependent pathophysiological changes in a rabbit model of high-fat diet-induced hyperlipidemia. FEBS Open Bio. 13(6): 1027-1040. doi: 10.1002/2211-5463.13597.
- Marangoni S, Ubiali M, Ambrosini F, Jahnel L, Vilani JM, Steagall PV and Vilani RGDC 2023. Effects of different rates of propofol with or without S-ketamine on ventricular function in healthy cats-a randomized study. Front Vet Sci. 11(10):1272949. doi: 10.3389/fvets.2023.1272949.
- Marques AEGW, Marques MG, Silveira BCR, Oliveira SP, Ferraz IG, Ventricci ABG, Silva NC, Nagata WB, Floriano BP, Ferreira WL and Santos PSP 2020. Lidocaine administered at a continuous rate infusion does not impair left ventricular systolic and diastolic function of healthy rabbits sedated with midazolam. Vet Anim Sci. 7(10): 100151. doi: 10.1016/j.vas.2020.100151.
- Orcutt CJ and Malakoff RL 2020. Cardiovascular Disease. In: Ferrets, rabbits, and rodents: clinical medicine and surgery. 4th ed. Quesenberry KE, Orcutt CJ, Mans C, Carpenter JW, (eds). Missouri: Elsevier. 250-257.
- Ozawa S, Sanchez-Migallon Guzman D, Keel K and Gunther-Harrington C 2021. Clinical and pathological findings in rabbits with cardiovascular disease: 59 cases (2001-2018). JAVMA. 259(7): 764-776. doi: 10.2460/javma.259.7.764.

- Pariaut R 2009. Cardiovascular physiology and diseases of the rabbit. Vet Clin North Am Exot Anim Pract. 12(1): 135-144, vii. doi: 10.1016/j.cvex.2008.08.004.
- Reusch B 2005. Investigation and management of cardiovascular disease in rabbits. In Pract. 27(8): 418-425. doi: 10.1136/inpract.27.8.418.
- Sahinovic MM, Struys MMRF and Absalom AR 2018. Clinical Pharmacokinetics and Pharmacodynamics of Propofol. Clin Pharmacokinet. 57(12): 1539-1558. doi: 10.1007/s40262-018-0672-3.
- Sández I, Verdier N, Redondo JI, Tarragona L, Donati PA, Serrano S and Otero PE 2022. Agreement between transthoracic echocardiography and esophageal Doppler on aortic flow variables in anesthetized mechanically ventilated dogs. Can Vet I. 63(7): 722-726.
- Schnellbacher R, Olson EE and Mayer J 2012. Emergency presentations associated with cardiovascular disease in exotic herbivores. J Exot Pet Med. 21(4): 316–327. doi: 10.1053/j.jepm.2012.09.007.
- Serres F, Chetboul V, Gouni V, Tissier R, Sampedrano CC and Pouchelon JL 2007. Diagnostic value of echo-Doppler and tissue Doppler imaging in dogs with pulmonary arterial hypertension. J Vet Intern Med. 21(6): 1280-1289. doi: 10.1892/07-064.1.
- Schober KE and Baade H 2006. Doppler echocardiographic prediction of pulmonary hypertension in West Highland White Terriers with chronic pulmonary disease. J Vet Intern Med. 20(4): 912-920. doi: 10.1892/0891-6640(2006)20[912:depoph]2.0.co;2.
- Szekeres R, Priksz D, Kiss R, Romanescu DD, Bombicz M, Varga B, Gesztelyi R, Szilagyi A, Takacs B, Tarjanyi V, Pelles-Tasko B, Forgacs I, Remenyik J, Szilvassy Z and Juhasz B 2023. Therapeutic Aspects of Prunus cerasus Extract in a Rabbit Model of Atherosclerosis-Associated Diastolic Dysfunction. International Journal of Molecular Sciences. 24(17): 13253. doi: 10.3390/ijms241713253.
- Turner Giannico A, Garcia DAA, Lima L, de Lara FA, Ponczek CAC, Shaw GC, Montiani-Ferreira F and Rodrigues Froes T 2015. Determination of normal echocardiographic, electrocardiographic, and radiographic cardiac parameters in the conscious New Zealand White rabbit. J Exotic Pet Med. 24(2): 223-234. doi: 10.1053/j.jepm.2015.04.013.
- Varga M 2014. Cardiorespiratory disease. In: Textbook of rabbit medicine. 2nd ed. M Varga (ed). New York: Elsevier. 390–404.
- Yang HS, Song BG, Kim JY, Kim SN and Kim TY 2013. Impact of propofol anesthesia induction on cardiac function in low-risk patients as measured by intraoperative Doppler tissue imaging. J Am Soc Echocardiogr. 26(7): 727-735. doi: 10.1016/j.echo.2013.03.016