Seroprevalence and associated risk factors of *Toxoplasma gondii* infections in dogs in Egypt

Mohamed Marzok^{1,2*} Hattan S. Gattan^{3,6} Omar A. AlJabr⁵ Mohammed H. Alruhaili^{4,6}
Mohamed Salem^{1,7} Abdelfattah Selim^{8*}

Abstract

Toxoplasma gondii is a parasite that is found all over the world and has significant importance in veterinary medicine and public health. Felids are considered a final host for the parasite and play an important role in the spreading of infection via contamination of the environment by oocysts. The study aimed to determine the seroprevalence of *T. gondii* and analyze risk factors associated with the infections in dogs. A total of 430 blood samples were collected from dogs from three governorates situated in northern Egypt and analyzed using commercial ELISA kit. The overall seroprevalence of *T. gondii* in dogs was 24%, and the Kafr ElSheikh governorate had the highest rate of seroprevalence at 29.3%. In addition, the seropositivity of *T. gondii* was higher in females (27.1%, 95 % CI: 22.28-32.42), dogs more 5 years age (32.6%, 95 % CI: 25.26-40.88), German Shepherd breed (31.5%, 95 % CI: 25.09-38.61), and in presence of rodents (31.5%, 95 % CI: 24.18-39.96). The multivariate logistic regression revealed that the sex, age, breed, and presence of rodents were potential risk factors for *T. gondii* infections in dogs. This study shows that *T. gondii* is a relevant parasite in Egypt, and more epidemiological surveys are needed to decrease the risk of human infection.

Keywords: Toxoplasma gondii, ELISA, Risk factors, Dogs, Egypt

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¹Department of Clinical Sciences, College of Veterinary Medicine, King Faisal University, Al-Ahsa 31982, Saudi Arabia

²Department of Surgery, Faculty of Veterinary Medicine, Kafr El Sheikh University, Kafr El Sheikh, Egypt.

³Department of Medical Laboratory Sciences, Faculty of Applied Medical Sciences, King Abdulaziz University, Jeddah, Saudi Arabia

⁴Department of Clinical Microbiology and Immunology, Faculty of Medicine, King AbdulAziz University, Jeddah, Saudi Arabia ⁵Department of Microbiology, College of Veterinary Medicine, King Faisal University, Al-Asha, P.O. Box 400, 31982, Saudi Arabia

⁶ Special Infectious Agents Unit, King Fahad Medical Research Center, King AbdulAziz University, Jeddah, Saudi Arabia ⁷Department of Medicine and Infectious Diseases, Faculty of Veterinary Medicine, Cairo University, 12613, Cairo, Egypt ⁸Department of Animal Medicine (Infectious Diseases), Faculty of Veterinary Medicine, Benha University, Toukh 13736, Egypt

^{*}Correspondence: Abdelfattah.selim@fvtm.bu.edu.eg, mmarzok@kfu.edu.sa (A. Selim, M. Marzok)

Introduction

Toxoplasma gondii, an intracellular obligate protozoan, is worldwide distributed and causes toxoplasmosis in all warm-blooded animals, including dogs and humans (Dubey 1987). The Centers for Disease Control (CDC) estimates that around one-third of people on the planet are infected with this parasite (Molan *et al.*, 2019).

The toxoplasma life cycle is quite intricate. *T. gondii* is a protozoan that forms cysts and is present in tissue. The life cycle of *T. gondii* consists of two stages: asexual and sexual. The sexual phase occurs in the intestines of definitive hosts (felids), whereas the asexual phase occurs in the tissues of intermediate hosts (Cenci-Goga *et al.,* 2011). The life cycle includes three infectious stages: bradyzoites, tachyzoites encased in tissue cysts, and sporozoites encapsulated in sporulated oocysts. *T. gondii* may be spread by different ways, including sexual transmission in a definite host and asexual transmission in intermediate hosts by meat-eating and vertical transmission (Dubey *et al.,* 2020; Smith *et al.,* 2021).

Human get *T. gondii* via eating undercooked or raw meat of lamb or pigs; however, environmental infection—for example, via oocysts in water, soil, or raw vegetables—may also contribute to the spread of the infection (Almeria and Dubey 2021; Kakakhel *et al.*, 2021).

Clinical toxoplasmosis in dogs is uncommon and typically associated with immunosuppression (Calero-Bernal and Gennari 2019). Some dogs were reported to have neuromuscular problems caused by T. gondii (Migliore et al., 2017; Patitucci et al., 1997). Furthermore, it has been reported that toxoplasmosis can cause and fatal respiratory distress severe immunocompromised dogs (Pepper et al., 2019). T. gondii infection is widespread; however, there have been few investigations into T. gondii in pets. The seroprevalence rates of T. gondii in dogs ranged from 28% to 98.0% (El Behairy et al., 2013; Khaled et al., 1982).

The diagnosis of toxoplasmosis infection is performed by isolation of the parasite from a sample of blood or feces, serological analysis, or using polymerase chain reaction (PCR) to identify specific DNA (Khan and Noordin, 2020; Reisberg *et al.*, 2013; Selim *et al.*, 2019; Selim *et al.*, 2013; Uddin *et al.*, 2021). Serological methods such as the enzyme-linked immunosorbent assay (ELISA), indirect fluorescent antibody test (IFAT), or other agglutination tests are often used to detect toxoplasmosis in humans and animals (Abbas *et al.*, 2020; Ybañez *et al.*, 2020).

Furthermore, data on *T. gondii* seroprevalence in dogs is still lacking for many Egyptian locations, and analysis of related risk factors for infection has yet to be done. Thus, the present study determined the seroprevalence of *T. gondii* in dogs from several governorates in Egypt. In addition, risk factors associated with the disease in dogs were also evaluated.

Materials and Methods

Ethical statement: This study was carried out in accordance with the regular rules and protocols established by the ethical committee of the Faculty of

Veterinary Medicine at Benha University, Egypt. Furthermore, the protocol and procedures of the study were approved by the ethical committee of the Faculty of Veterinary Medicine, Benha University. Blood samples were obtained by highly experienced veterinarians after the animal owners verbally agreed to participate in the study. Throughout the investigation, the ARRIVE criteria were followed.

Study areas: The samples were collected from animals living in three governorates located in Northern Egypt from January to December 2022. These governorates include Kafr ElSheikh, Gharbia, and Giza, as shown in Figure 1.

Kafr ElSheikh Governorate is recognized for its desert climate, with an average annual temperature of 24°C and negligible rainfall throughout the year. Gharbia has a subtropical desert climate (Classification: BWh), which is located at an elevation of 8.49 meters above sea level, and the annual temperature in the city is 24°C. The summer in Giza is long, humid, hot, arid, and clear, while the winter is cold, dry, and mainly clear, and the temperature ranges between 25-45°C throughout the year.

Sample size and sampling: The following formula was used to calculate the required sample size (Thrusfield 2018).

$$n = Z^2 P(1 - P)/d^2$$

Where n is the sample size, Z is the confidence level, P is the previously reported prevalence rate (23.3%) for *T. gondii* in dogs in Egypt by Salama *et al.* (2022), and d is the accuracy. A total of 430 blood samples were collected from the cephalic veins of randomly selected dogs using vacutainer tubes. The samples were centrifuged at 3500 rpm for 10 to separate sera and kept at 20°C till serological analysis.

Questionnaire: Data of examined animals were collected during sampling from owners *via* a previously prepared questionnaire. Animal characteristics such as age (>2, 2-5, >5 years), breed (German Shepherd, Rottweiler, and Pitbull), sex (male and female), area (rural and urban), and presence of rodents in the house were all gathered.

Serological analysis: Dog serum samples were analyzed for anti-T. gondii antibodies using commercial ELISA kits (ID Screen® Toxoplasmosis Indirect Multi-species, IDVet, Grables, France). The kits included both positive and negative control sera, and the test was carried out in accordance with the manufacturer's guidelines. The ELISA reader (AMR 100, AllSheng, China) was used to measure the optical density (OD) of the findings at 450 nm.

The Toxoplasmosis kit uses a peroxidase-conjugated anti-multi-species secondary antibody to identify particular immunoglobulin G (IgG) antibodies against the P30 *T. gondii* protein. The following formula was used to determine the percentage of sample (S) to positive (P) ratio (S/P%):

S/P%= OD sample - OD negative control/ OD positive control - OD negative control × 100

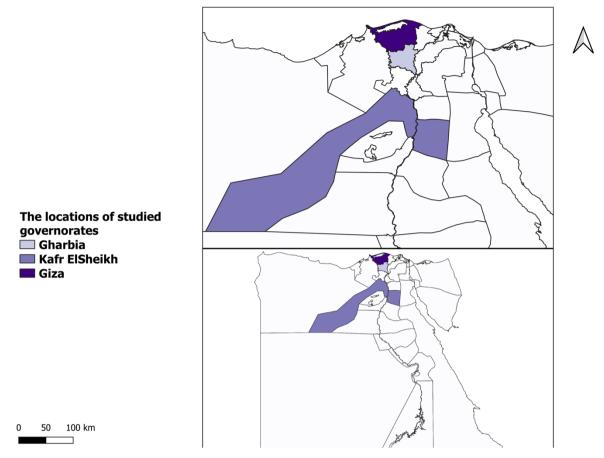


Figure 1 The locations of studied governorates (MAP generated using QGiS software)

 Table 1
 Seroprevalence of T. gondii in dogs in relation to different factors

| Variable | Total examined animal | No of positive | No of negative | % of positive | 95%CI | Statistic | |
|--------------------|-----------------------------|----------------|-------------------|---------------|-------------|------------------------------------|--|
| Locality | | | | | | | |
| Kafr ElSheikh | 150 | 44 | 106 | 29.3 | 22.63-37.06 | | |
| Gharbia | 140 | 25 | 115 | 17.9 | 12.4-25.04 | χ2=5.248 df=2 P=0.073 | |
| Giza | 140 | 34 | 106 | 24.3 | 17.94-32.02 | P=0.073 | |
| Sex | | | | | | | |
| Male | 138 | 24 | 114 | 17.4 | 11.97-24.57 | χ 2=4.804 df=1 | |
| Female | 292 | 79 | 213 | 27.1 | 22.28-32.42 | P=0.028* | |
| Age | | | 0 | | | | |
| <2 | 110 | 16 | 94 | 14.5 | 9.16-22.33 | 2-10.027 | |
| 2-5 | 185 | 43 | 142 | 23.2 | 17.74-29.83 | χ2=10.927 df=2 <i>P</i> =0.004* | |
| >5 | 135 | 44 | 91 | 32.6 | 25.26-40.88 | u1=2 P=0.004° | |
| Breed | | | | | | | |
| German Shepherd | 178 | 56 | 122 | 31.5 | 25.09-38.61 | χ2=11.073 | |
| Rottweiler | 132 | 29 | 103 | 22.0 | 15.75-29.77 | df=2 P=0.004* | |
| Pitbull | 120 | 18 | 102 | 15.0 | 9.7-22.47 | u_1 -2 P -0.004 | |
| Area | | | | | | | |
| rural | 180 | 30 | 150 | 16.7 | 11.93-22.8 | χ 2=9.025 df=1 | |
| Urban | 250 | 73 | 177 | 29.2 | 23.91-35.12 | P=0.003* | |
| Presence of rodent | | | | | | | |
| Yes | 130 | 41 | 89 | 31.5 | 24.18-39.96 | χ 2=5.885 df=1 | |
| No | 300 | 62 | 238 | 20.7 | 16.47-25.61 | P=0.015* | |
| Total | 430 | 103 | 327 | 24.0 | 20.16-28.2 | _ | |

^{*}Results considered significant if the *P* value is less than 0.05

| Variables | В | S.E. | OR | 95% CI for OR | P value |
|--------------------|-------|-------|------|---------------|---------|
| Sex | | J.L. | OK | 3370 CITOLOR | 1 varae |
| Female | 0.549 | 0.270 | 1.73 | 1.02-2.94 | 0.042 |
| Age | | | | | |
| Age 2-5 | 0.434 | 0.332 | 1.54 | 0.81-2.96 | 0.192 |
| >5 | 0.844 | 0.339 | 2.33 | 1.19-4.52 | 0.013 |
| Breed | | | | | |
| German Shepherd | 0.751 | 0.314 | 2.12 | 1.15-3.92 | 0.017 |
| Rottweiler | 0.291 | 0.342 | 1.34 | 0.68-2.61 | 0.394 |
| Area | | | | | |
| Urban | 0.694 | 0.251 | 2.00 | 1.22-3.27 | 0.006 |
| Presence of rodent | | | | | |
| Yes | 0.497 | 0.249 | 1.64 | 1.01-2.68 | 0.046 |

 Table 2
 Multivariate analysis for risk factors associated with T. gondii infection in dogs

B: Logistic regression coefficient, SE: Standard error, OR: Odds ratio, CI: Confidence interval

According to the manufacturer, samples with S/P% values more than 50% were regarded as positive, those between 40 and 50% were regarded as dubious, and measures less than or equal to 40% were regarded as negative.

Result

In the three governorates under study, the overall seroprevalence of *T. gondii* in dogs was 24%, with no significant variation between the governorates. The seroprevalence was higher in Kafer ElSheikh (29.3%) than in Giza (24.3%) and Gharbia (17.9%), Table 1.

The results of the univariate analysis showed a significant association (P<0.05) between seroprevalence of T. gondii and sex, age, breed, area, and presence of rodents. Females (27.1%, 95 % CI: 22.28-32.42) and German Shepherd breed (31.5 %, 95 % CI: 25.09-38.61) had a significantly higher seroprevalence of T. gondii. Furthermore, dogs raised in urban areas or in close contact with rodents had a higher T. gondii seroprevalence of 29.2% and 31.5%, respectively.

The results of the multivariate logistic regression model revealed that the following factors were found to be associated with an increased risk of *T. gondii* infection in dogs: female (OR=1.73, 95 % CI: 1.02-2.94), dogs older than 5 years (OR=2.33, 95 % CI: 1.19-4.52), German Shepherd breed (OR=2.12, 95 % CI: 1.15-3.92), raising dogs in an urban area (OR=2, 95 % CI: 1.22-3.27), and contact with rodents (OR=1.64, 95 % CI: 1.01-2.68), Table 2.

Discussion

The prevalence study of *T. gondii* in felids is always very necessary to reduce oocyst adulteration in the environment and to transmit infections to humans (Marzok *et al.*, 2023; Selim *et al.*, 2023a; Selim *et al.*, 2023b). Serological detection is a reliable and accurate approach to identifying *T. gondii* infection in dogs because clinical could not be a specific way to identify the infection. *T. gondii* infection may be diagnosed by examining various tissues; however, this requires the animals to be dead (Ali *et al.*, 2003; Björkman *et al.*, 1994; Dubey *et al.*, 2020). In the present study, a serosurvey for *T. gondii* infection in dogs was conducted to determine the seroprevalence of antibodies against *T. gondii* and to assess the associated risk factors.

The seroprevalence in this study was non significantly varied between studied governorates, and the overall seroprevalence of *T. gondii* in dogs was 24%, which comes in agreement with the previously reported rate of 23.3% in Egypt by Salama *et al.* (2022). The seroprevalence rate of this study was higher than that reported in Iran (8.94%) (Hosseininejad and Hosseini 2011) and Taiwan (7.9%) (Lin 1998). In other studies, the higher seroprevalence for *T. gondii* in dogs was observed, it was 45.1% in Brazil (Azevedo *et al.*, 2005), 30% in Sweden (Björkman *et al.*, 1994), 51.3% in Turkey (Oncel *et al.*, 2007), 67% in Senegal (Ahmad *et al.*, 2001), 39% in Lahore (Shahzad *et al.*, 2006) and 50% in Vietnam (Dubey *et al.*, 2007).

It is possible to attribute this variation in seroprevalence between countries to environmental and geographical factors, serological tests used, the diversity of cultures and behaviors, socioeconomic conditions, and sanitary conditions, dog populations, as well as parasite intrinsic factors like genetic diversity (Liu *et al.*, 2012; Marques *et al.*, 2019; Rodrigues *et al.*, 2016; Selim and Abdelhady 2020; Selim *et al.*, 2021a; Selim *et al.*, 2021e).

Contrary to the findings of Azevedo *et al.* (2005), sex was significantly associated with *T. gondii* seropositivity in dogs, and it was higher in females than males. The current results agreed with the findings of Arunvipas *et al.* (2013) but differed from those of other studies (Ali *et al.*, 2003; Cañón-Franco *et al.*, 2004). In contrast, Salama *et al.* (2022) found that males, especially the German Shepherd breed, had higher seropositivity for *T. gondii* when compared to females and other breeds.

The high seroprevalence seen in dogs older than five years may be explained by the increased chance of parasite interaction with age (Ahmad *et al.*, 2014). Elderly dogs showed higher seroprevalence, which comes in agreement with previously reported findings by Wu *et al.* (2011) and Pena *et al.* (2006). Adult dogs are more likely to be exposed to *T. gondii* oocysts due to their larger wandering behavior and potential interaction with contaminated surroundings, waste, and leftover meals (Selim *et al.*, 2021b; Watanabe *et al.*, 2020).

The current study revealed that the breed was substantially associated with *T. gondii* seropositivity, and the German Shepherd is the most vulnerable breed, in contrast to the findings of Raimundo *et al.* (2015), Yan *et al.* (2012), and Azevedo *et al.* (2005).

Based on present findings, *T. gondii* seropositivity increased significantly among urban dogs than rural ones, which comes in accordance with Saldanha-Elias *et al.* (2019) and Watanabe *et al.* (2020). The risk of *T. gondii* may be increased by various urban-specific characteristics such as a high cat population, human activity resulting in more garbage cans on the street, and exposure to undercooked meat. (Fábrega *et al.*, 2020).

Notably and consistent with our results, prior research revealed a positive association between toxoplasmosis and the presence of rodents (Silva *et al.*, 2010). This might be explained by the fact that rodents are nocturnal creatures that may not have been seen despite being present in the home and drawing in cats.

In conclusion, *T. gondii* infection exists in dogs, according to the assessment of serological data and their association with the examined epidemiological factors. The results of this study revealed that sex, age, breed, area, and presence of rodents are significant risks for *T. gondii* infection. This study also highlights the necessity of looking at dogs' involvement in toxoplasmosis cases as part of public health monitoring initiatives due to the shared sources of infection between people and dogs.

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Authors' contributions: Conceptualization, methodology, formal analysis, investigation, resources, data curation, writing-original draft preparation, A.S., H.S.G., M.H.A., M.S., M.M., O.A.A. and A.A.; writing-review and editing, A.S., M.H.A., M.S., H.S.G., O.A.A. and M.M..; project administration, M.M.; funding acquisition, A.S., M.H.A., M.S., H.S.G., O.A.A. and M.M. All authors have read and agreed to the published version of the manuscript.

References

- Abbas IE, Villena I and Dubey JP 2020. A review on toxoplasmosis in humans and animals from Egypt. Parasitology. 147: 135-159.
- Ahmad F, Maqbool A, Mahfooz A and Hayat S 2001. Serological survey of *Toxoplasma gondii* in dogs and cats. Pak Vet J. 21: 31-35.
- Ahmad N, Ahmed H, Irum S and Qayyum M 2014. Seroprevalence of IgG and IgM antibodies and associated risk factors for toxoplasmosis in cats and dogs from sub-tropical arid parts of Pakistan. Trop Biomed. 31(4): 777-784.
- Ali C, Harris JA, Watkins JD and Adesiyun AA 2003. Seroepidemiology of *Toxoplasma gondii* in dogs in Trinidad and Tobago. Vet Parasitol. 113 (3-4): 179-187.
- Almeria S and Dubey JP 2021. Foodborne transmission of *Toxoplasma gondii* infection in the last decade. An overview. Res Vet Sci. 135: 371-385.
- Arunvipas P, Jittapalapong S, Inpankaew T, Pinyopanuwat N, Chimnoi W and Maruyama S 2013. Seroprevalence and risk factors influenced transmission of *Toxoplasma gondii* in dogs and cats in dairy farms in Western Thailand. Afr J Agric Res. 8: 591-595.
- Azevedo SS, Batista CS, Vasconcellos SA, Aguiar DM, Ragozo AM, Rodrigues AA, Alves CJ and Gennari SM 2005. Seroepidemiology of *Toxoplasma gondii* and *Neospora caninum* in dogs from the state of Paraíba, Northeast region of Brazil. Res Vet Sci. 79: 51-56.
- Björkman C, Lundén A and Uggla A 1994. Prevalence of Antibodies to *Neospora caninum* and *Toxoplasma gondii* in Swedish Dogs. Acta Vet Scand. 35: 445 – 447.
- Calero-Bernal R and Gennari SM 2019. Clinical Toxoplasmosis in Dogs and Cats: An Update. Front Vet Sci. 6: 54.
- Cañón-Franco WA, Bergamaschi DP, Labruna MB, Camargo LMA, Silva JCS, Pinter A and Gennari SM 2004. Occurrence of Anti-*Toxoplasma gondii* Antibodies in Dogs in the Urban Area of Monte Negro, Rondônia, Brazil. Vet Res Commun. 28: 113-118.
- Cenci-Goga BT, Rossitto PV, Sechi P, McCrindle CM and Cullor JS 2011. Toxoplasma in animals, food, and humans: an old parasite of new concern. Foodborne Pathog Dis. 8: 751-762.
- Dubey JP 1987. Toxoplasmosis. Vet Clin North Am Small Anim Pract. 17: 1389-1404.
- Dubey JP, Huong LT, Sundar N and Su C 2007. Genetic characterization of *Toxoplasma gondii* isolates in dogs from Vietnam suggests their South American origin. Vet Parasitol. 146: 347-351.

- Dubey JP, Murata FHA, Cerqueira-Cézar CK, Kwok OCH, Yang Y and Su C 2020. *Toxoplasma gondii* infections in dogs: 2009-2020. Vet Parasitol. 287: 109223.
- El Behairy AM, Choudhary S, Ferreira LR, Kwok OC, Hilali M, Su C and Dubey JP 2013. Genetic characterization of viable *Toxoplasma gondii* isolates from stray dogs from Giza, Egypt. Vet Parasitol. 193: 25-29.
- Fábrega L, Restrepo CM, Torres AdS, Smith D, Chan P, Pérez D, Cumbrera A and Caballero EZ 2020. Frequency of *Toxoplasma gondii* and Risk Factors Associated with the Infection in Stray Dogs and Cats of Panama. Microorganisms. 8.
- Hosseininejad M and Hosseini F 2011. Seroprevalence of *Neospora caninum* and *Toxoplasma gondii* infection in dogs from west and central parts of Iran using two indirect ELISA tests and assessment of associate risk factors. Iran J Vet Res. 12: 46-51.
- Kakakhel MA, Wu F, Anwar Z, Saif I, Akbar NU, Gul N, Ali I, Feng H and Wang W 2021. The presence of *Toxoplasma gondii* in soil, their transmission, and their influence on the small ruminants and human population: A review. Microb Pathog. 158: 104850.
- Khaled ML, Morsy TA, Sadek MS and Salama MM 1982. The presence of antibodies against toxoplasmosis, leishmaniasis and amoebiasis in stray dogs in Cairo, Egypt. J Egypt Soc Parasitol. 12: 341-347.
- Khan AH and Noordin R 2020. Serological and molecular rapid diagnostic tests for Toxoplasma infection in humans and animals. Eur J Clin Microbiol Infect Dis. 39: 19-30.
- Lin DS 1998. Seroprevalences to *Toxoplasma gondii* in privately-owned dogs in Taiwan. Prev Vet Med. 35: 21-27.
- Liu Y, He G, Cheng Z, Qi Y, Liu J, Zhang H, Liu G, Shi D, Yang D, Wang S and Wang Z 2012. Seroprevalence of *Toxoplasma gondii* in Dogs in Shandong, Henan, and Heilongjiang Provinces, and in the Xinjiang Uygur Autonomous Region, People's Republic of China. J Parasitol. 98: 211-212
- Marques SB, Caldart ET, Palavro ACF, Ferreira FP, Cordi A and Pasquali AKS 2019. Seroprevalence of *Toxoplasma gondii* in dogs and cats domiciled in the west of Santa Catarina, Brazil. Iniciação Científica Cesumar. 21(2):115.
- Marzok M, Al-Jabr OA, Salem M, Alkashif K, Sayed-Ahmed M, Wakid MH, Kandeel M and Selim A 2023. Seroprevalence and Risk Factors for *Toxoplasma gondii* Infection in Horses. Vet Sci. 10(3): 237.
- Migliore S, La Marca S, Stabile C, Di Marco Lo Presti V and Vitale M 2017. A rare case of acute toxoplasmosis in a stray dog due to infection of *T. gondii* clonal type I: public health concern in urban settings with stray animals?. BMC Vet Res. 13: 249
- Molan A, Nosaka K, Hunter M and Wang W 2019. Global status of *Toxoplasma gondii* infection: systematic review and prevalence snapshots. Trop Biomed. 36: 898-925.

- Oncel T, Handemir E, Kamburgil K and Yurtalan S 2007. Determination of seropositivity for *Toxoplasma gondii* in stray dogs in Istanbul, Turkey. Revue De Medecine Veterinaire. 158: 223-228
- Patitucci AN, Alley MR, Jones BR and Charleston WA 1997. Protozoal encephalomyelitis of dogs involving *Neospora caninum* and *Toxoplasma gondii* in New Zealand. N Z Vet J. 45: 231-235.
- Pena HFJ, Soares RM, Amaku M, Dubey JP and Gennari SM 2006. *Toxoplasma gondii* infection in cats from São Paulo state, Brazil: seroprevalence, oocyst shedding, isolation in mice, and biologic and molecular characterization. Res Vet Sci. 81 (1): 58-67.
- Pepper A, Mansfield C, Stent A and Johnstone T 2019. Toxoplasmosis as a cause of life-threatening respiratory distress in a dog receiving immunosuppressive therapy. Clin Case Rep. 7: 942-948.
- Raimundo JM, Guimarães A, Moraes LMdB, Santos LA, Nepomuceno LL, Barbosa SM, Pires MS, Santos HA, Massard CL, Machado RZ and Baldani CD 2015. *Toxoplasma gondii* and *Neospora caninum* in dogs from the state of Tocantins: serology and associated factors. Revista brasileira de parasitologia veterinaria. 24 (4): 475-481.
- Reisberg K, Selim AM and Gaede W 2013. Simultaneous detection of *Chlamydia* spp., *Coxiella burnetii*, and *Neospora caninum* in abortion material of ruminants by multiplex real-time polymerase chain reaction. J Vet Diagn Invest. 25: 614-619.
- Rodrigues JY, Almeida AdBPFd, Boa Sorte EdC, Gasparetto ND, Cruz FACSd and Sousa VRF 2016. Seroprevalence of *Toxoplasma gondii* in dogs of riverside communities of Mato Grosso Pantanal, Brazil. Rev Bras Parasitol Vet. 25(4): 531-535.
- Salama DB, Fereig RM, Abdelbaky HH, Shahat MS, Arafa WM, Aboelhadid SM, Mohamed AE, Metwally S, Abas O and Suo X 2022. *Toxoplasma gondii* and *Neospora caninum* antibodies in dogs and cats from Egypt and risk factor analysis. Pathogens. 11: 1464.
- Saldanha-Elias AM, Silva MA, Silva VO, Amorim SLA, Coutinho AR, Santos HA, Giunchetti RC, Vitor RWA and Geiger SM 2019. Prevalence of Endoparasites in Urban Stray Dogs from Brazil Diagnosed with Leishmania, with Potential for Human Zoonoses. Acta Parasitol. 64: 352 359.
- Selim A and Abdelhady A 2020. Neosporosis among Egyptian camels and its associated risk factors. Trop Anim Health Prod. 52: 3381-3385.
- Selim A, Abdelhady A and Alahadeb J 2021a. Prevalence and first molecular characterization of *Ehrlichia canis* in Egyptian dogs. Pak Vet J. 41(1): 117-121.
- Selim A, Abdelrahman A, Thiéry R and Sidi-Boumedine K 2019. Molecular typing of *Coxiella* burnetii from sheep in Egypt. Comp Immunol Microbiol Infect Dis. 67: 101353.
- Selim A, Alanazi AD, Sazmand A and Otranto D 2021b. Seroprevalence and associated risk factors for

- vector-borne pathogens in dogs from Egypt. Parasit Vectors. 14: 1-11.
- Selim A, Alshammari A, Gattan HS, Alruhaili MH, Rashed GA and Shoulah S 2023a. Seroprevalence and associated risk factors for *Toxoplasma gondii* in water buffaloes (*Bubalus bubalis*) in Egypt. Comp Immunol Microbiol Infect Dis. 101: 102058.
- Selim A, Attia KA, Alsubki RA, Kimiko I and Sayed-Ahmed MZ 2022. Cross-sectional survey on *Mycobacterium avium* Subsp. paratuberculosis in Dromedary Camels: Seroprevalence and risk factors, Acta Trop. 226: 106261.
- Selim A, El-Haig M, Galila ES and Gaede W 2013. Direct detection of *Mycobacterium avium* subsp. Paratuberculosis in bovine milk by multiplex Real-time PCR. Anim Sci Pap Rep. 31: 291-302.
- Selim A, Khater H and Almohammed HI 2021c. A recent update about seroprevalence of ovine neosporosis in Northern Egypt and its associated risk factors. Sci Rep. 11: 14043.
- Selim A, Manaa EA, Alanazi AD and Alyousif MS 2021d. Seroprevalence, risk factors and molecular identification of bovine leukemia virus in Egyptian cattle. Animals. 11: 319.
- Selim A, Marzok M, Alshammari A, Al-Jabr OA, Salem M and Wakid MH 2023b. *Toxoplasma gondii* infection in Egyptian domestic sheep and goats: seroprevalence and risk factors. Trop Anim Health Prod. 55: 182.
- Selim A, Shoulah S, Abdelhady A, Alouffi A, Alraey Y and Al-Salem WS 2021e. Seroprevalence and risk factors associated with canine leishmaniasis in Egypt. Vet Sci. 8: 236.
- Shahzad A, Khan MS, Ashraf K, Avais MA, Pervez K and Khan Ja 2006. Sero-epidemiological and haematological studies on toxoplasmosis in cats, dogs and their owners in Lahore, Pakistan. J Protozool Res. 16: 60-73.
- Silva RCd, Souza LCd, Langoni H, Tanaka E, Lima VYd and Silva AVd 2010. Risk factors and presence of antibodies to *Toxoplasma gondii* in dogs from the coast of São Paulo State, Brazil. Pesqui Vet Bras. 30: 161-166.
- Smith NC, Goulart C, Hayward JA, Kupz A, Miller CM and van Dooren GG 2021. Control of human toxoplasmosis. Int J Parasitol. 51: 95-121.
- Thrusfield M 2018. Veterinary epidemiology, (John Wiley & Sons).
- Uddin A, Hossain D, Ahsan MI, Atikuzzaman M and Karim MR 2021. Review on diagnosis and molecular characterization of *Toxoplasma gondii* in humans and animals. Trop Biomed. 38: 511-539.
- Watanabe M, Sadiq MB, Mulop NIA, Mohammed K, Rani PAM, Fong LS, Aziz NA, Kamaludeen J, Ramanoon SZ, Mansor R, Ping TL and Syed-Hussain SS 2020. Prevalence of *Toxoplasma gondii* Antibodies in Stray Dogs from Various Locations in West and East Malaysia, Korean. J Parasitol. 58: 487-492.
- Wu S-M, Huang S-Y, Fu B, Liu G, Chen J-x, Chen M-x, Yuan Z-G, Zhou DH, Weng Y-B, Zhu X-Q and Ye D-H 2011. Seroprevalence of *Toxoplasma gondii* infection in pet dogs in Lanzhou, Northwest China. Parasit Vectors. 4: 64 64.

- Yan C, Fu L-l, Yue C-l, Tang R-x, Liu Y-s, Lv L, Shi N, Zeng P, Zhang P, Wang D-H, Zhou DH, Zhu X-Q and Zheng K-Y 2012. Stray dogs as indicators of *Toxoplasma gondii* distributed in the environment: the first report across an urban-rural gradient in China. Parasit Vectors. 5: 5 5.
- Ybañez RHD, Ybañez AP and Nishikawa Y 2020. Review on the Current Trends of Toxoplasmosis Serodiagnosis in Humans. Front Cell Infect Microbiol. 10: 204.