# Yield, composition, fatty acid profile and CLA content of milk from goats fed with different levels of OPF

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#### Abstract

The objective of this study was to evaluate the effect of different inclusion levels of OPF in total mixed ration (TMR)based diet on yield, composition, FA profile and CLA content of milk in dairy goats. Six nulliparous female dairy goats (Saanen x Thai native, 19-21 months old, first lactation) were randomly set according to a 3x3 Latin square design with 2 replications for receiving diets consisted of 0% OPF (OPF0), 20% OPF (OPF20) and 40% OPF (OPF40). The average weight of the goats was 32.48±1.30 kg (mean±s.d.). The findings of this study showed that dry matter intake (DMI) and all nutrient intake of the goats increased significantly as OPF inclusion was increased (p<0.05), excluding crude protein intake (CPI) (p>0.05). Goats from OPF40 group had the highest CPI, however, there was no significant difference (p>0.05) in CPI of OPF20 and OPF0 groups. Feeding OPF altered the composition of protein and lactose, density and freezing point (p<0.05). However, it did not influence (p>0.05) the composition of fat, solid non-fat and minerals in the milk. There were no significant differences in oleic acid, dihomo-γ-linoleic acid, and eicosapentaenoic acid in milk fat of goats (p>0.05). 40% OPF inclusion increased the production of LnA, DHA, CLA and the ratio of PUFA:SFA; and lowered AA production and n-6:n-3 ratio of goat milk (*p*>0.05). It can be concluded that 40% OPF inclusion in TMRbased diet can be used as an alternative to producing goat's milk with enriched potential health benefits.

Keywords: oil palm fronds, TMR-based diet, dairy goats, milk fatty acids profile, CLA

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## Introduction

In several countries, goat's milk has become one of the economic interests and an alternative for those who have allergies to dairy cow products (Schettino et al., 2017). Goat's milk can be categorized as a functional food due to the large number of bioactive components and the easiness of digestion (Savoini et al., 2019). Nutritional quality in food products has become important because there is an increase in consumer awareness of the link between health and diet (Hilali et al., 2018). In the human diet, milk and dairy products are the major sources of saturated fatty acids (SFAs) (Bayat et al., 2018). It is well documented that humans with high SFA consumption tend to have a higher risk of cardiovascular disease (CVD). Conversely, dietary polyunsaturated fatty acids (PUFAs) from dairy products has been associated with many health benefits (Yurchenko et al., 2018; Markey et al., 2017; Hilali et al., 2018). PUFAs, in particular conjugated linoleic acid (CLA), has physiological effects as an antimutagenic, anti-tumor, anti-diabetic (type II), an agent of body fat reduction (anti-obesity), anti-carcinogenic (fight against cancer), an agent of atherosclerosis prevention, an immune function enhancer, an antihypertensive, a hyperglycemia reduction agent and bone mineralization improvement agent in human body (Koba and Yanagita, 2013; Bouattour et al., 2008; Gomez-Cortes et al., 2018; Tudisco et al., 2015; Castro et al., 2009; Jacobs et al., 2011).

Although the composition of fatty acid (FA) in milk is affected by biohydrogenation of rumen and conversion of Δ9-desaturase enzyme, modifying animals diet can make large changes to the milk FA profile (Liu et al., 2016; Yurchenko et al., 2018; Bayat et al., 2018; Hilali et al., 2018). Basal diet composition together with source and type of lipid supplementation can affect milk FA composition (Bayat et al., 2018). Animals fed with fresh pasture tend to have a better CLA and vaccenic acid (VA) contents in milk compared to animals fed with dried forages. The possible reason is either fresh pasture may improve the growth of rumen's specific bacteria that is liable to CLA production or inhibit the VA to stearic acid final reduction (Nudda et al., 2005). Moreover, dietary PUFA sources decreased medium-chain SFA and n-6:n-3 ratio and increased milk CLA concentration, which are favorable as milk with a better nutritional quality for human consumption (Morsy et al., 2015; Marin et al., 2011; Castro et al., 2009).

Oil palm frond (OPF) is a fibrous crop residue that is abundantly available (Dalton et al., 2017). OPF has been utilized as a long-term ruminant feed with no toxic effect (Hassan et al., 1991). OPF contains PUFAs, especially linolenic acid (LnA; C18:3n-3) and linoleic acid (LA; C18:2n-6) (Hassim et al., 2010). The inclusion of OPF in a ruminant diet can increase the proportion of unsaturated fatty acid (UFA) in sheep plasma and rumen contents via regulation of rumen biohydrogenation or restriction of microbial access to facilitate the continuous availability of dietary UFA (Ghani et al., 2017). Nevertheless, information regarding dietary different levels of OPF to improve yield, composition, FA profile and CLA content of milk in dairy goats is limited. Therefore, the objective of this study was to provide a report on different OPF inclusion levels in TMR-based diet to produce goat's milk with an improved FA profile and CLA content.

#### Materials and Methods

Animals, Housing and Experimental Diets: Six lactating dairy goats (Saanen x Thai native, 19-21 months old, first lactation) were blocked by parity and balanced for milking days and body weight. The animals were then randomly arranged in a 3x3 Latin square design with 2 replications. The average weight of the goats was 32.48±1.30 kg (mean±s.d.). All animal handling procedures and experimental protocols were based on the Institutional Animal Care and Use Committee, Prince of Songkla University (U1-01633-2558), based on the Ethical Principles and Guidelines for the Use of Animals, National Research Council of Thailand. Each goat was kept in a stainless steel individual pen in the goat section of the Animal Innovation Production and Management Division, Faculty of Natural Resources, Prince of Songkla University, Hat Yai Campus. Ivermectin injection (Ivomec) was administered to provide efficacy against parasitic infestations.

The goats were provided free access to water and mineral blocks. Three treatment diets consisting of a basal diet (OPF0; 40% rice straw + 60% concentrate), the basal diet with 50% rice straw replacement with OPF (OPF20; 20% rice straw + 20% OPF + 60% concentrate) and the basal diet with 100% rice straw replacement with OPF (OPF40; 40% OPF + 60% rice straw). All treatment diets were set up as isonitrogenous and iso-caloric total mixed rations (TMR). The diets were provided ad libitum. A Tenera hybrid breed (Dura x Pisifera) more than 5 years old from the Faculty of Natural Resources, Prince of Songkla University was used and was harvested daily. Rice straw and OPF were chopped into 0.5-2 cm length and then mixed with grains on a daily basis to comply with the treatment diets.

The experiment was carried out in three periods. Each period was divided into 14 days for adaptation time and 30 days for data collection. Body weights (BW) of the goats were recorded weekly before morning feeding. Feed intake and milk production were recorded daily throughout the data collection.

Data Collection and Laboratory Analysis Procedures: Feed Sampling Procedures: The samples of diets were collected once every 5 days to analyze feed composition. Additional feed samples were taken to determine the FA composition. The standard methods of AOAC (1998) were used to analyse the dry matter (DM), organic matter (OM), crude protein (CP), ether extract (EE) and ash of the feed and residual feed. Neutral detergent fiber (NDF) and acid detergent fiber (ADF) were analyzed according to the method described by Van Soest et al. (1991) with adaptation for Fiber Analyzer. The amount of EE was obtained by extracting the samples with dichloromethane using a Soxtec System based on AOAC (1998). All nutrient compositions were expressed based on the final DM. A calorimeter (LECO AC500) was used to determine the gross energy (GE) of the diets.

Milk Yield, Milk Composition and Milk FA Profile: Milk samples were collected twice a day (120 ml each), in the morning and afternoon milking during the data collection time. An additional milk sample from each goat (120 ml) was collected for milk FA profile analysis. Milk samples were stored at -20°C until milk composition and milk FA analysis. The samples of milk were sent to a laboratory for fat, protein, total solid, solid non-fat and lactose analysis (AOAC, 1990) by mid-infrared spectrophotometry (Lactostar).

Lipid in milk samples was extracted following the method of Zhang et al. (2015). The samples of milk from each goat were pooled and then extracted in 4 mL of isopropanol and hexane (ratio 2/3, v/v), then followed by the solution of 2 mL sodium sulfate and centrifuged at 2,500 x g at 20°C for 20 mins. The supernatant was moved into a hydrolysis tube and dried with a nitrogen flow. A 2 mL of NaOCH<sub>3</sub>-methanol was then pipetted to the dry sample. The mixture was entirely blended using a vortex mixer and heated for 15 mins at 50°C. After the period of cooling, 2 mL of methanol was added to the mixture and provide a time of 15 mins at 80°C for the mixture to react. After cooling down, 3 mL of deionized water and 6 mL of hexane were added to was completely mixture. The mixture homogenized utilizing a vortex mixture and was centrifuged at 2,500 x g at 20°C for 20 mins. The supernatant was poured into a graduated tube with hexane and fitted to 10 mL. Approximately 1 g of anhydrous sodium sulfate was added.

Fatty acid methyl esters (FAMEs) of feed and milk samples were analyzed using gas chromatography (GC) fitted with an HP-88 fused-silica capillary column (100 m x 0.25 mm with a film thickness of 0.2  $\mu$ m). The temperature of the column was initially held at 120°C for 10 mins. Afterward, it was heated up to 230°C at 3.2°C/mins and held for 35 mins. The injector

temperature was kept at  $250^{\circ}$ C and the detector temperature was maintained at  $300^{\circ}$ C. The volume of injection was 1  $\mu$ L. Each peak was identified by utilizing the pure methyl ester standards.

Statistical Analysis: Analysis of variance (ANOVA) for 3x3 Latin Square using SPSS Statistics version 16 was performed on the DM, nutrient intake, milk yield, milk composition and milk FA profile to study the influence of the treatment diets. The parametric model was denoted as  $X_{ijk} = \mu + \alpha_i + \beta_j + \gamma_k + \varepsilon_{ijk}$ , where  $X_{ijk}$  is the examination from animal j with diet i, in period k. Whereas,  $\mu$  stands for the overall mean,  $\alpha_i$  is the OPF levels effect (i = 0%, 20%, 40%),  $\beta_j$  is the animal effect (j = 1, 2, 3, 4, 5, 6),  $\gamma_k$  is the period effect (k = 1, 2, 3) and  $\varepsilon_{ijk}$  is the residual effect. Duncan's new multiple range tests were further performed to determine differences between treatment means (Steel and Torrie, 1980.

### Results

Chemical Compositions and Fatty Acids Profile of Treatment Diets: The chemical compositions and fatty acids (FA) profile of the treatment diets are presented in Table 1 and Table 2. The inclusion of OPF increased the moisture content and lowered the ash content of TMR-based diets. All treatment diets had similar CP and gross energy (GE) contents. The treatment diet with the highest percentages of EE and fiber was OPF40, followed by OPF20 and OPF0. OPF40 and OPF20 had a similar amount of LnA, whereas OPF0 was the lowest. Compared to the diet without OPF inclusion, OPF20 and OPF40 contained a higher FA from the n-3 family. A similar PUFA:SFA ratio was indicated in all treatment diets. OPF inclusion in TMRbased diet lowered n-6:n-3 ratio. The ratio of n-6:n-3 in OPF0, OPF20, and OPF40 were 8.7:1, 5.1:1, and 4.8:1, respectively.

 Table 1
 Chemical compositions of experimental diets

Items	Treatment <sup>1(</sup>				
itenis	OPF0	OPF20	OPF40		
Ingredients (%)					
OPF	0.00	20.00	40.00		
Rice straw	40.00	20.00	0.00		
Soybean meal	25.15	23.01	21.66		
Corn	28.86	30.99	32.34		
Molasses	3.00	3.00	3.00		
Premix	3.00	3.00	3.00		
Chemical compositions					
DM (%)	87.82	67.04	55.62		
CP %)DM(	17.86	17.43	17.07		
EE %)DM(	1.80	2.42	2.84		
Ash %)DM(	10.44	8 <b>.</b> 65	7 <b>.</b> 57		
NDF %)DM(	31.61	38.07	48.80		
ADF %)DM(	19.31	20.26	22.04		
Gross energy )Cal/kg DM(	3984.93	4021.32	4087.62		

DM:dry matter; CP:crude protein; EE:ether extract; NDF:neutral digestible fiber; ADF:acid detergent fiber.

<sup>&</sup>lt;sup>1)</sup> OPF0: 60% concentrate + 40% rice straw; OPF20: 60% concentrate + 20% rice straw + 20% oil palm fronds; OPF40: 60% concentrate + 40% oil palm fronds.

Table 2 Fatty acids profile of experimental diets

Itama	Treatment <sup>1(</sup>				
Items	OPF0	OPF20	OPF40		
Fatty acids profile )g/100 g FA(2(					
C18:0, Stearic acid	1.61	2.48	1.81		
C18:1n9c, Oleic acid	8.21	10.22	9.82		
C18:1n9t, Elaidic acid	0.20	0.22	0.30		
C18:2n6c, Linoleic acid (LA)	16.67	20.78	16.59		
C18:2n6t, Linolelaidic acid	1.85	1.09	1.35		
C18:3n3, α-Linolenic acid (LnA)	2.08	5.10	5.06		
C18:3n6, γ-Linolenic acid	7.99	6.68	8.54		
C20:0, Eicosanoic acid	0.41	0.46	0.22		
C20:2, Eicosadienoic acid	0.32	0.11	0.11		
C20:3n3, Eicosatrienoic acid	0.44	0.11	0.09		
C20:4n6, Arachidonic acid) AA(	0.18	0.09	0.09		
C22:0, Docosanoic acid	0.33	0.36	0.23		
C22:2, Cetoleic acid	0.56	0.12	0.12		
C24:0, Tetracosanoic acid	0.73	0.47	0.44		
SFA	58.29	54.35	56.07		
MUFA	10.55	11.08	11.52		
PUFA	31.16	34.56	32.41		
Total n-3 <sup>3(</sup>	3.43	5.65	5.62		
Total n-6 <sup>4(</sup>	26.84	28.68	26.56		
PUFA:SFA	0.54	0.64	0.58		
n-6:n-3	8.67	5.12	4.78		

<sup>&</sup>lt;sup>1</sup> OPF0 :60 %concentrate +40 %rice straw; OPF20 :60 %concentrate +20 %rice straw +20 %oil palm fronds; OPF40 :60 %concentrate +40 %oil palm fronds.

Dry Matter and Nutrient Intake: The dry matter and nutrient intake of dairy goats fed with different inclusion levels of OPF are shown in Table 3. The findings of this study showed that dry matter intake (DMI) and all nutrients intake of the goats increased significantly as OPF inclusion increased (p<0.05), except for crude protein intake (CPI). Goats from the OPF40 group showed the highest CPI, however, there was no significant difference (p>0.050 in CPI of OPF20 and OPF0 groups. Based on percent body weight (%BW), the DMI of goats fed with OPF40 was 1.14 and 1.06 fold higher than those fed with OPF0 and OPF20, respectively. Fat, fiber and OM intake of the goats increased as the inclusion of OPF was increased. The intakes of EE, OM, NDF, and ADF of the OPF40 group were the highest.

Milk Yield and Milk Composition: Milk yield and milk composition of dairy goats fed with OPF in TMR based diet are shown in Table 4. The highest milk yield was reported in the OPF40 group. However, there was no significant difference in the milk yield of OPF0 and OPF20 groups. Although OPF0 and OPF20 had similar milk yield, goats from OPF20 group produced higher nutrient due to its higher milk density. Goats with higher inclusion of OPF produced higher fat, solid nonfat (SNF), protein, lactose and minerals per day. The highest production of those nutrients was on the OPF40 group, while the lowest was on the OPF0 group. The inclusion of OPF in TMR-based diet of dairy goats altered the composition of milk protein, lactose, density and freezing point but did not alter the composition of milk fat, solid non-fat, and minerals.

Table 3 Dry matter and nutrients intake of lactating dairy goats fed with different levels of oil palm fronds

Item		Treatment <sup>1)</sup>			D1
	OPF0	OPF20	OPF40	SEM	P-value
DM Intake					
Total (g/d)	1113.3°	1170.1 <sup>b</sup>	1292.6a	13.5	< 0.001
%BW	$3.40^{c}$	3.63b	3.86a	0.06	< 0.001
g/kg BW <sup>0.75</sup>	80.9c	86.5b	92.3a	1.46	< 0.001
Nutrient intake (g/d)					
OMI	996.8°	1068.9b	1194.7a	12.4	< 0.001
CPI	198.8ь	203.9b	220.6a	2.32	< 0.001
EEI	$20.0^{\circ}$	28.3b	36.7a	0.36	< 0.001
NDFI	351.8c	445.4 <sup>b</sup>	630.8a	6.12	< 0.001
ADFI	214.9°	237.1ь	284.9a	2.89	< 0.001

SEM :standard error of the means) n =3(; DMI :dry matter intake; BW :body weight; CPI :crude protein intake; EEI :ether extract intake; NDFI :neutral digestible fiber intake; ADFI :acid detergent fiber intake; OMI :organic matter intake.

<sup>&</sup>lt;sup>2</sup> (FA :fatty acids; SFA :sum of saturated fatty acids; MUFA :sum of monounsaturated fatty acids; PUFA :sum of polyunsaturated fatty acids.

<sup>&</sup>lt;sup>3</sup> (Sum of n-6 fatty acids.

<sup>4)</sup> Sum of n-3 fatty acids.

<sup>&</sup>lt;sup>1)</sup> OPF0: 60% concentrate + 40% rice straw; OPF20: 60% concentrate + 20% rice straw + 20% oil palm fronds; OPF40: 60% concentrate + 40% oil palm fronds.

 $_{a,b,c}$  Means in the same row with different superscript are statistically different (p<0.05).

Table 4 Milk yield and milk composition of dairy goats fed with oil palm frond in TMR based diet.

Items	T	Treatment <sup>1)</sup>			D1
	OPF0	OPF20	OPF40	SEM	P-value
Yield					
Milk (kg/d)	1.08b	1.09b	1.40a	17.2	< 0.01
Fat (g/d)	$30.90^{\circ}$	34.20b	41.80a	0.54	< 0.01
Solid non-fat (g/d)	88.15c	93.71 <sup>b</sup>	116.89a	1.41	< 0.01
Protein (g/d)	31.81°	33.86 <sup>b</sup>	42.03a	0.51	< 0.01
Lactose (g/d)	46.82c	48.99b	62.10a	0.76	< 0.01
Minerals (g/d)	10.41°	10.93 <sup>b</sup>	13.57a	0.18	< 0.05
Composition					
Fat (%)	2.87	3.03	3.00	0.05	2.62
Solid non-fat (%)	8.18	8.65	8.33	0.15	0.12
Protein (%)	2.95 <sup>b</sup>	$3.14^{a}$	$3.00^{b}$	0.02	< 0.01
Lactose (%)	4.35c	4.62a	4.43b	0.02	< 0.01
Minerals (%)	0.97	0.98	0.97	0.01	0.09
Density	1.0244 <sup>c</sup>	1.0259a	$1.0248^{b}$	0.01	< 0.01
Freezing point (°C)	-0.5541 <sup>b</sup>	-0.5761a	-0.5557a	0.003	< 0.01

SEM :standard error of the means) n =6(; OPF0 :60 %concentrate +40 %rice straw; OPF20 :60 %concentrate +20 %rice straw +20 % oil palm fronds; OPF40 :60 %concentrate +40 %oil palm fronds.

Milk Fatty Acid Profile: Table 5 presents the milk fatty acid profile of dairy goats fed with different inclusion levels of OPF. OPF inclusion up to 40% in the treatment significantly increased (p < 0.05)concentrations of elaidic acid (C18:1n9t), linolelaidic acid (C18:2n6t) and α-linolenic acid (ALA; C18:3n3). Similar concentrations of stearic acid (C18:0), eicosatrienoic acid (C20:3n3), and docosahexaenoic acid (DHA; C22:6n3) concentrations were indicated in OPF20 and OPF40 groups (p>0.05). Meanwhile, OPF0 had lower concentrations of those FAs (p<0.05). There was no significant difference in linoleic acid (LA; C18:2n6c) of OPF20 and OPF40 groups. However, the values were lower than the OPF0 group. The lowest concentration of arachidonic acid (AA; C20:4n6) was in the diet with 40% OPF inclusion. Dietary OPF did not alter oleic acid (C18:1n9c), dihomo-y-linoleic acid (DGLA; C20:3n6), and eicosapentaenoic acid (EPA; C20:5n3) in milk fat of goats (p>0.05). OPF40 contained the highest PUFA and CLA contents and the highest UFA:SFA ratio compared to OPF20 and OPF0 (p<0.05). There was no significant difference in SFA concentration and n-6:n-3 ratio of OPF40 and OPF20, however, the values were significantly lowered than OPF 0. The ratio between n-6 and n-3 in OPF40 and OPF20 was 3.66 and 4.11, respectively.

### Discussion

Chemical Compositions and Fatty Acid Profile of Treatment Diets: All treatment diets were formulated as iso-nitrogen and iso-caloric diets, so they were similar in CP and GE contents. Similar values for CP and EE of diets with OPF inclusion were previously reported by Ebrahimi et al. (2015), and Hamchara et al. (2018), respectively. The higher moisture content of OPF20 and OPF40 could be attributed to the form of OPF used in this study that was fresh and was chopped daily. NDF and ADF contents of OPF20 and OPF40 were also close to the value reported by Ebrahimi et al. (2015). Previous studies also reported that linolenic acid (C18:3 n-3) in OPF (24.97%) is higher than in rice straw (1.72%) (Ebrahimi et al., 2015; Taira, 1983).

Therefore, the higher the inclusion of OPF in the diet of the present study, the lower the ratio of n-6:n-3 FA.

Dry Matter and Nutrient Intake: The DMI of OPFO, OPF20, and OPF40 were 3.40%, 3.63%, and 3.86% BW, respectively. Exotic breeds raised in tropical areas have typical DMI values of about 3.6% BW (Stares et al., 1992). It means that OPFO had DMI lower than the suggested DMI for Saanen in tropical areas. The findings of this study also indicate that higher moisture content, low palatability of rice straw and lower n-6:n-3 ratio due to OPF inclusion significantly increased DMI and nutrient intake of lactating dairy goats, although diets with higher OPF inclusion contained higher levels of NDF. The treatment diets with OPF inclusion had 60-70% DM, while the diet without OPF contained 90% DM. Adequate moisture content in complete feed can increase palatability and DMI by improving texture and diluting undesirable flavors (Lahr et al., 1983). Similar to the findings of this study, Meiske and Goodrich (1971) also reported that feeding feedlot steers with a diet containing roughly 66% DM resulted in the maximum DMI. Moreover, high silica content in rice straw also contributes to its lower palatability and thus feed intake (Nguyen et al., 2020; Oladosu et al., 2016).

We did not observe the effect of n-6:n-3 ratios on fermentation patterns, however, lowered n-6:n-3 ratios in the diet with OPF inclusion may have changed rumen microbial population, thus it could overcome the effect of NDF on DMI. Decreased level of n-6:n-3 PUFA ratios in the diet increases the proportion of cellulolytic bacteria namely R. albus and R. flavefaciens (Ebrahimi et al., 2017). Additionally, Greco et al. (2015), showed a consistent result with this study that feeding a 3.9:1 ratio of n-6 and n-3 FA resulted in lactating Holstein cows with better DMI. PUFAs potentially affect hepatic oxidation and have the ability to stimulate the release of gut peptides to alter DMI. Opposite results have been reported where dietary different n-6:n-3 ratio did not influence the DMI of lactating goats and growing lambs (Bouattour et al., 2008; Kim et al., 2007).

a,b,c Means in the same row with different superscript are statistically different (p<0.05).

Milk Yield and Milk Composition: Higher milk production of OPF40 compared to OPF0 and OPF20 was the result of greater nutrient intake, especially CPI. Protein is an essential limiting nutrient for producing milk and achieving high yields in ruminants (Salo, 2018). Based on Looper (1994), feed provides the nutrients that are the precursors, either directly or indirectly, of the principal milk solids. Thus, an increase in feed intake usually results in the production of a greater volume of milk.

There was no significant difference in milk fat percentage amongst the treatment diets. However, the maintains the rumen pH (Banakar *et al.*, 2018).

milk fat percentages of OPF20 and OPOF40 were between the range of Saneen goats during early lactation to late lactation (2.99 to 3.66) as reported by Kljajevic *et al.* (2018), while the OPF0 group was lower than the suggested range. This could be due to higher fiber intake (NDF and ADF) of goats fed with OPF. One of the factors that may affect fat production of dairy animals is the fiber content of the feed. Increasing fiber content and physically effective NDF enhances saliva flow, acetate to propionate ratio, milk fat levels and

Table 5 Milk fatty acids profile of lactating dairy goats fed with oil palm frond in TMR based diet.

•	Treatment <sup>1)</sup>				
Items	OPF0	OPF20	OPF40	SEM	P-value
Fatty acids profile (g/100 g FA) <sup>2)</sup>					
C4:0, Butyric acid	1.52c	1.97 <sup>b</sup>	2.42a	0.135	0.003
C6:0, Caproic acid	1.47	1.60	1.62	0.089	0.445
C8:0. Caprylic acid	0.71 <sup>c</sup>	$0.98^{b}$	1.15a	0.034	0.000
C10:0, Capric acid	1.49	1.49	1.39	0.052	0.296
C12:0, Lauric acid	1.83a	$1.64^{b}$	$1.37^{\circ}$	0.035	0.000
C14:0, Myristic acid	7.85a	7.43 <sup>b</sup>	6.81 <sup>c</sup>	0.102	0.000
C14:1, Myristoleic acid	$0.74^{a}$	$0.67^{b}$	0.67 <sup>b</sup>	0.01	0.001
C15:0, Pentadecanoic acid	0.97a	$0.86^{b}$	$0.82^{b}$	0.02	0.001
C16:0, Palmitic acid	27.72a	25.79b	23.79c	0.308	0.000
C16:1, Palmitoleic acid	1.45	1.45	1.39	0.03	0.209
C17:0, Heptadecanoic acid	0.62a	0.60a	$0.46^{b}$	0.02	0.000
C18:0, Stearic acid	12.08b	12.29a	12.26a	0.049	0.025
C18:1n9t, Elaidic acid	6.20c	7.49 <sup>b</sup>	9.20a	0.244	0.000
C18:1n9c, Oleic acid	29.54	29.12	29.47	0.203	0.306
C18:2n6t, Linolelaidic acid	$0.54^{c}$	0.71 <sup>b</sup>	1.01a	0.031	0.000
C18:2n6c, Linoleic acid (LA)	2.25a	2.06b	2.00b	0.051	0.016
C18:3n3, α-Linolenic acid (LnA)	$0.58^{c}$	$0.91^{b}$	1.18a	0.044	0.000
C18:3n6, γ-Linolenic acid	0.01	0.01	0.01	0.003	0.927
C20:0, Eicosanoic acid	0.21a	0.19a	$0.14^{b}$	0.009	0.001
C20:1, Gadoleic acid	0.06a	$0.03^{b}$	$0.02^{b}$	0.005	0.001
C20:2, Eicosadienoic acid	$0.10^{a}$	0.09a	0.05ь	0.01	0.025
C20:3n3, Eicosatrienoic acid	$0.01^{b}$	0.02a	0.02a	0.002	0.004
C20:3n6, Dihomo-γ-linoleic acid (DGLA)	0.02	0.02	0.03	0.004	0.513
C20:4n6, Arachidonic acid (AA)	0.16a	0.15a	0.13b	0.007	0.014
C20:5n3, Eicosapentaenoic acid (EPA)	0.04	0.04	0.05	0.005	0.669
C22:0, Docosanoic acid	0.11	0.45	0.23	0.136	0.260
C22:1n9, Erucic acid	0.02	0.02	0.02	0.004	0.784
C22:2, Cetoleic acid	0.02	0.01	0.01	0.004	0.955
C22:6n3, Docosahexaenoic acid (DHA)	0.03b	0.12a	0.13a	0.012	0.000
C23:0, Tricosanoic acid	0.03a	0.02ab	0.01b	0.006	0.095
C24:0, Tetracosanoic acid	0.03	0.02	0.03	0.006	0.452
C9,T11	1.17 <sup>b</sup>	1.27b	1.63a	0.056	0.000
C9,C11	0.02	0.02	0.02	0.003	0.919
T9,T11	$0.12^{c}$	$0.15^{b}$	0.18a	0.006	0.000
SFA	56.90a	55.65a	52.80b	0.417	0.000
MUFA	38.05 <sup>b</sup>	38.78 <sup>b</sup>	40.76a	0.323	0.000
PUFA	5.06c	5.58b	6.44a	0.108	0.000
n-6 <sup>3)</sup>	4.30b	4.39b	5.01a	0.079	0.000
n-3 <sup>4)</sup>	0.65 <sup>c</sup>	1.08 <sup>b</sup>	1.37a	0.044	0.000
n6:n3	7.16a	4.11 <sup>b</sup>	3.66b	0.233	0.000
PUFA:SFA	0.09b	0.10b	0.12a	0.003	0.000

<sup>&</sup>lt;sup>1</sup> OPF0 :60 %concentrate +40 %rice straw; OPF20 :60 %concentrate +20 %rice straw +20 %oil palm fronds; OPF40 :60 %concentrate +40 %oil palm fronds.

In the present study, there were divergences in the milk protein and lactose compositions among dairy goats fed with different levels of OPF inclusion. One explanation for these results may be related to the relative amounts of energy and protein available in the rumen. Previous studies have addressed the fact that milk protein content may respond to energy-protein interaction, although the results have been varied

<sup>&</sup>lt;sup>2</sup> (FA :fatty acids; SFA :sum of saturated fatty acids; MUFA :sum of monounsaturated fatty acids; PUFA :sum of polyunsaturated fatty acids.

<sup>&</sup>lt;sup>3</sup> (Sum of n-6 fatty acids.

<sup>4)</sup> Sum of n-3 fatty acids.

(Brun-Lafleur et al., 2010). Furthermore, a tangible candidate for the relationship between protein supply and lactose output is glucose, in which the whole body (WB) rate of appearance (Ra) of glucose (total of real gluconeogenesis, glycogenolysis and portal absorption) increases with increasing protein supply via casein infusion (Lapierre et al., 2010).

Milk Fatty Acid Profile: The milk FA profile of goats in this study was dominated by oleic acid, palmitic acid, stearic acid, elaidic acid and myristic acid. The findings were in agreement with studies by Ferrand-Calmels et al. (2014), Cossignani et al. (2014), and Lopez et al. (2019). However, the presence of caproic acid (C6:0), caprylic acid (C8:0) and capric acid (C10:0) in the present study were lower than the reported values of those previous studies. Caproic, caprylic and capric acids are medium-chain FAs that have been named after goats because of their prevalence in goat's milk. They makeup to 15% of total FA in goat's milk is responsible for its flavor (Kompan and Komprej, 2012). Dietary high-level pasture tends to lower the production omf capric acid in goat's milk (LeDoux et al., 2002). Goats fed with OPF40 contained the highest caprylic acid which can give beneficial effects. Caprylic acid is one of the alternatives as intra-mammary infusion against bovine mastitis (Nair at al., 2005) and is effective in inactivating infant pathogens such as respiratory syncytial virus and herpes simplex virus (Isaacs et al., 1995).

Feeding OPF40 to lactating dairy goats resulted in milk with lower AA and higher DHA contents, compared to OPF0. AA is categorized as a PUFA essential for normal health. However, it also contributes to the inflammation process and leads to the promotion of mediators responsible for wound healing and resolving inflammation (Tallima and Ridi, 2017). On the other side, DHA has been known for playing an important role in inflammation, immunity and the functional development and growth of the infant's brain (Lee and Jenkins, 2011; Horrocks and Yeo, 1999). DHA is a biohydrogenation product of LnA. LnA turns into stearidonic acid (18:4n3) and eicosatetraenoic acid (20:4)n-3) eicosapentaenoic acid (EPA; 20:5n3). In its later stage, EPA is metabolized to be eicosanoids or DHA (Schmitz and Ecker, 2008). Goats fed with OPF20 and OPF40 yielded similar DHA in milk fat due to similar amounts of LnA in those diets. Both diets also contained LnA higher than OPF0, therefore, a significantly higher amount of DHA was indicated in milk fat of OPF20 and OPF40 groups.

The lower amount of SFA in OPF40 was the consequence of a higher intake of fiber. A linear result was reported by Hassim *et al.* (2010), that higher OPF inclusion rates decreased SFA production in the fermentation pattern of in vitro incubations due to the increasing amount of fiber. In the present study, goats fed with OPF40 received the highest amount of fresh forage. Therefore, it may have led to the highest CLA production. Several factors can affect the variation of CLA contained in milk, such as forage types, forage conservation methods, breeds and lactation stage (Halmemies-Beauchet-Filleau *et al.*, 2016; Bouattour *et al.*, 2008). Previous studies mentioned that modifying

an animal's diet could increase the amount of CLA in milk (Hur *et al.*, 2007). Nudda *et al.* (2005), reported that animals fed with fresh pasture tended to have better CLA content in milk compared to animals fed with dried forages. The possible reason is either fresh pasture may improve the growth of rumen's specific bacteria that is liable on CLA production or inhibit the VA to stearic acid final reduction. Additionally, dietary fresh grass speeded up CLA synthesis via stearoyl-CoA desaturase activation (Hur *et al.*, 2017).

The n-6:n-3 ratios of OPF20 and OPF40 were 4.11 and 3.66, respectively. Those values were within the range of recommendation (1:1 to 4:1) for human consumption to achieve normal development and homeostasis (Simopoulos, 2008). Finally, milk from the OPF40 group is likely to be preferable due to its higher PUFA:SFA ratio. Recently, consumers have been advised to limit their SFA intake and replace SFA with unsaturated fat (Kris-Etherthon and Krauss, 2020).

The inclusion of OPF enhances the DMI and milk yield of lactating dairy goats. Feeding OPF to lactating dairy goats alters the composition of protein and lactose, density and the freezing point of milk. However, it did not affect the percentage of fat, solid non-fat and minerals in milk. The inclusion of 40% OPF in TMR-based diet increased the production of LnA, DHA, CLA, and PUFA:SFA ratio. It also decreased AA production and n-6:n-3 ratio of goat milk. Feeding 40% OPF in TMR-based diet can be used as an alternative to producing goat milk with enriched potential health benefits.

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## References

AOAC 1990. Official Methods of Analyses. 15th Ed. Virginia: Association of Official Analytical Chemists Inc.

AOAC 1998. Official Methods of Analysis of AOAC International. 16<sup>th</sup> ed. Gaithersburg: AOAC International.

Bayat AR, Tapio I, Vikki J, Shingfield KJ, Leskinen H 2018. Plant oil supplements reduce methane emissions and improve milk fatty acid composition in dairy cows fed grass silage-based diets without affecting milk yield. J Dairy Sci. 101:1136-1151.

- Banakar PS, Anand KN, Shashak CG, Lakhani N 2018. Physically effective fibre in ruminant nutrition: A review. J Pharmacog Phytochem. 7(4): 303-308.
- Bouattour MA, Csals R, Elbanell E, Such X, Caja G 2008. Feeding soybean oil to dairy goats increases conjugated linoleic acid in milk. J Dairy Sci. 91: 2399-2407.
- Brun-Lafleur L, Delaby L, Husson F, Faverdin P 2010. Predicting energy x protein interaction on milk yield and milk composition in dairy cows. J Dairy Sci. 93(9): 4128-4143.
- Castro T, Manso T, Jimeno V, Alamo MD, Mantecon AR 2009. Effects of dietary sources of vegetable fats on performance of dairy ewes and conjugated linoleic acid (CLA) in milk. Small Rumin Res. 84:47-53
- Cossignani L, Giua L, Urbani E, Simonetti MS, Blasi F 2014. Fatty acid composition and CLA content in goat milk and cheese samples from Umbrian market. Eur Food Res Technol. 239: 905-911. doi: 10.1007/s00217-014-2287-8
- Dalton OS, Mohamed AF, Chikere AO 2017. Status evaluation of palm oil waste management sustainability in Malaysia. OIDA Int J Sustain Dev. 10(12): 41-47.
- Ebrahimi M, Rajion MA, Jafari S, Jahromi MF, Oskoueian E, Sazili AQ, Goh YM, Ghaffari MH 2019. Effects of dietary *n*-6: *n*-3 polyunsaturated fatty acid ratios on meat quality, carcass characteristics, tissue fatty acid profiles, and expression of lipogenic genes in growing goats. PLoS One. 14(9): 1-21. doi:10.1371/journal. pone.0188369
- Ebrahimi M, Rajion MA, Adeyemi KD, Jafari S, Jahromi MF, Oskoueian E, Goh YM, Ghaffari MH 2017. Dietary n-6:n-3 Fatty Acid Ratios Alter Rumen Fermentation Parameters and Microbial Populations in Goats. J Agric Food Chem. 65(4): 737-744. doi: 10.1021/acs.jafc.6b04704
- Ebrahimi M, Rajion MA, Meng GY, Shokryzadan P, Sazili AQ, Jahroni MF 2015. Feeding oil Palm (Elaeis guineensis, Jacq.) fronds alter rumen protozoal population and ruminal fermentation pattern in goats. Ital J Anim Sci. 14(3): 403-409. doi: 10.4081/ijas.2015.3877
- Ferrand-Calmels M, Palhiere I, Brochard M, Leray O, Astruc JM, Aurel MR, Barbey S, Bouvier F, Brunschwig P, Caillat H, Douguet M, Faucon-Lahalle F, Gele M, Thomas G, Trommenschlager JM, Larroque H 2014. Prediction of fatty acid profiles in cow, ewe, and goat milk by mid-infrared spectometry. J Dairy Sci. 97(1): 17-35. doi: 10.3168/jds.2013-6648
- Ghani AAA, Rusli ND, Shahudin MS, Goh YM, Zamri-Saad M, Hafandi A, Hassim HA 2017. Utilisation of oil palm fronds as ruminant feed and its effect on fatty acid metabolism. Pertanika J Tropic Agric Sci. 40: 215-224.
- Gomez-Cortez P, Juarez M, de la Fuente MA 2018. An update version: Milk fatty acids and potential health benefits. Trend Food Sci Technol. 81: 1-9.
- Greco LF, Neto JTN, Pedrico A, Ferrazza RA, Lima FS, Bisinotto RS, Martinez N, Garcia M, Ribeiro

- ES, Gomes GC, Shin JH, Ballou MA, Thatcher WW, Staples CR, Santos JE 2015. Effects of altering the ratio of dietary n-6 to n-3 fatty acids on performance and inflammatory responses to a lipopolysaccharide challenge in lactating Holstein cows. J Dairy Sci. 98(1): 602-617. doi:10.3168/jds.2014-8805
- Halmemies-Beauchet-Filleau A, Shingfield KJ, Simpura I, Kokkonen T, Jaakkola S, Toivonen V, Vanhatalo A 2016. Article in Press: Effect of incremental amounts of camelina oil on milk fatty acid composition in lactating cows fed diets based on a mixture of grass and red clover silage and concentrates containing camelina expeller. J Dairy Sci. 100: 1-20.
- Hassan OA, Oshio S, Ismail AR, Jaafar DM, Nakanishi N, Dahlan I, Ong SH 1991. Experiences and challenges in processing, treatment, storage and feeding of oil palm trunks based dietsfor beef production. Proceedings of Seminar on Oil Palm Trunks and Other Palmwood Utilization. 4-5 March 1991, Kuala Lumpur, Malaysia. Pp. 231-245.
- Hassim HA, Lourenço M, Goel G, Vlaeminck B, Goh YM, Fievez V 2010. Effect of different inclusion levels of oil palm fronds on in vitro rumen fermentation pattern, fatty acid metabolism and apparent biohydrogenation of linoleic and linolenic acid. Anim Feed Sci Technol. 162(3): 155-158. doi:10.1016/j.anifeedsci.2010.09.011
- Hegde MV, Zanwar AA, Adekar SP. 2016. Omega-3 Fatty Acids: Keys to Nutritional Health. Switzerland: Springer International Publishing.
- Hilali M, Risxhkowsky B, Iniguez L, Mayer H, Schreiner M 2018. Changes in the milk fatty acid profile of Awassi sheep in response to supplementation with agro-industrial by-products. Small Rumin Res. 166: 93-100.
- Horrocks LA, Yeo YK 1999. Health benefits of docosahexaenoic acid (DHA). Pharmacol Res. 40(3): 211-225. doi: 10.1006/phrs.1999.0495
- Hur SJ, Park GB, Joo ST 2007. Biological activities of conjugated linoleic acid (CLA) and effects of CLA in animal products. Livest Sci. 110: 221-229.
- Isaacs CE, Litov RE, Thormar H 1995. Antimicrobial activity of lipids added to human milk, infant formula, and bovine milk. Biochem. 6(7): 362-366. Doi: 10.1016/0955-2863(95)80003-U
- Jacobs AAA, Baal JV, Smits MA, Taweel HZH, Hendriks WH, Vuuren AMV, Dijkstra J 2011. Effects of feeding rapeseed oil, soybean oil, or linseed oil on strearoyl-CoA desaturase expression in the mammary gland of dairy cows. J Dairy Sci. 94: 874-887.
- Kljajevic NV, Tomasevic IB, Miloradovic ZN, Nedeljkovic A, Miocinovic JB, Jovanovic ST 2018. Seasonal variations of Saanen goat milk composition and the impact of climatic conditions. J Food Sci Technol. 55(1): 299-303. doi:10.1007/s13197-017-2938-4
- Kim SC, Adesogan AT, Badinga L, Staples CR 2007. Effects of dietary n-6:n-3 fatty acid ratio on feed intake, digestibility and, fatty acid profiles of the

- ruminal contents, liver and, muscle of growing lambs. J Anim Sci. 85:706–716.
- Koba K, Yanagita T 2013. Health benefits of conjugated linoleic acid (CLA), Obes Res Clin Pract. 8(6): 525-532.
- Kompran D, Komprej A 2012. The effect of fatty acids in goat milk on health. Milk Production: An Up-To Date Overview of Animal Nutrition, Management and Health. INTECH. doi: 10.5772/50769
- Kris-Etherthon PM, Krauss RM 2020. Public health guidelines should recommend reducing saturated fat consumption as much as possible: yes. Am J Clin Nutr. 00: 1-6. Doi: 10.1093/ajcn/nqaa110
- Lahr AD, Otterby DE, Johnson DG, Linn JG, Lundquist RG 1983. Effects of moisture content of complete diets on feed intake and milk production by cows. J Dairy Sci. 66(9): 1891-1900.
- Lapierre H, Galindo CE, Lemosquet S, Marty IO, Doepel L, Ouellet DR 2010. Protein supply, glucose kinetics and milk yield in dairy cows. Proceedings of the 3<sup>rd</sup> EAAP International Smposium on Energy and Protein Metabolism and Nutrition (ISEP), Oct 2010, Parme, Italy. pp. 275-286.
- LeDoux M, Rouzeau A, Bas P, Saivantt D 2002.

  Occurence of trans-C18:1 fatty acid isomers in goat milk: effect of two dietary regimens. J Dairy Sci. 85(1):190-197. doi:10.3168/jds.s0022-0302(02)74067-8
- Lee YJ, Jenkins TC 2011. Identification of enriched conjugated linoleic acid isomers in cultures of ruminal microorganisms after dosing with 1
  13C-linoleic acid. J Microbiol. 49: 622-627.
- Liu C, Li D, Chen W, Li Y, Wu H, Meng Q, Zhou Z 2016. Estimating ruminal crude protein degradation from beef cattle feedstuff. Sci Rep. 9: 11368. doi.org/10.1038/s41598-019-47768-3
- Looper M 1994. Factors Affecting Milk Composition of
  Lactating Cows. Division of Agriculture
  Research & Extension FSA4014. University of
  Arkansas. https://www.uaex.edu/
  publications/pdf/FSA-4014.pdf [accessed on
  April 29, 2020 at 8:16PM]
- Lopez A, Vasconi M, Moretti VM, Bellagamba F 2019. Fatty acid profile in goat milk from high- and low-input conventional and organic system. Animals (Basel). 9(7): 452. Doi: 10.3390/ani9070452.
- Marin ALM, Gomez-Cortez P, Castro AGG, Juarez M, Alba LMP, Hernandez MP, Fuente MADL 2011.

  Animal performance and milk fatty acid profile of dairy goats fed diets with different unsaturated plant oils. J Dairy Sci. 94: 5359-5368.
- Markey O, Souroullas K, Fagan CC, Kliem KE, Vasilopoulou D, Jackson KG, Humphries DJ, Grandison AS, Givens DI, Lovegrove JA, Methven L 2017. Consumer acceptance of dairy products with a saturated fatty acid-reduced, monounsaturated fatty acid-enriched content. J Dairy Sci. 100: 7953-7966.
- Meiske JC, Goodrich RD 1971. Backgrounding feedlot cattle. Minnesota Nutr Conf Proc. 15.

- Morsy TA, Kholif SM, Kholif AE, Matloup OH, Salem AZM, Elella AA 2015. Influence of sunflower whole seeds or oil on ruminal fermentation, milk production, composition and fatty acid profile in lactating goats. Asian-Australas J Anim Sci. 28: 1116–1122.
- Nair MKM, Joy J, Vasudevan P, Hinckley L, Hoagland TA, Venkitanarayanan KS 2005. Antibacterial effect of caprylic acid and monocaprylin on major bacterial mastitis pathogens. J Dairy Sci. 88(10): 3488-3495. doi: 10.3186/jds.S0022-0302(05)73033-2
- Nguyen DV, Vu CC, Nguyen TV 2020. The Current Utilisation and Possible Treatments of Rice Straw as Ruminant Feed in Vietnam: A Review. Pak J Nutr. 19: 91-104. doi: 10.3923/pjn.2020.91.104
- Nudda A, McGuire MA, Battacone G, Pulina G 2005. Seasonal variation in conjugated linoleic acid and vaccenic acid in milk fat of sheep and its transfer to cheese and ricotta. J Dairy Sci. 88(4): 1311–1319.
- Oladosu Y, Rafii MY, Abdullah N, Magaji U, Ramli GHA, Miah G 2016. Fermentation Quality and Additives: A Case of Rice Straw Silage. BioMed Res Int. 2016. doi: 10.1155/2016/7985167
- Salo, S 2018. Effects of quality and amounts of dietary protein on dairy cattle reproduction and the environment. Dairy and Vet Sci J. 5(5): 1-7. doi: 10.19080/JDVS.2018.05.555675
- Savoini G, Zorini FO, Farina G, Agazzi A, Cattaneo D, Invernizzi G 2019. Effects of fat supplementation in dairy goats on lipid metabolism and heath status. Animals (Basel). 9(11): 917. doi: 10.3390/ani9110917
- Schettino B, Vega S, Gutierrez R, Escobar A, Romero J, Dominguez E 2017. Fatty acid profile of goat milk in diets supplemented with chia seed (*Salvia hispanica L.*). J Dairy Sci. 100: 1-10.
- Schmitz G, Ecker J 2008. A Review: The opposing effects of n-3 and n-6 fatty acids. Prog Lipid Res. 47: 147-155
- Simopoulos AP 2008. The importance of the omega-6/omega-3 fatty acid ratio in cardiovascular disease and other chronic diseases. Exp Biol Med. 233: 674-688. doi: 10.3181/0711-MR-311
- Stares JES, Said AN, Kategile JA 1992. The complementarity of feed resources for animal production in Africa. Proceedings of the Joint Feed Resources Networks Workshop. 4-8 March 1991, Gaborone, Bostwana. African Feeds Research Network. ILCA (International Livestock Center for Africa), Addis Ababa, Ethiopia. pp. 430.
- Steel RGD, Torrie JH 1980. Principal and Procedures of Statistics: A Biometrical Approach. 2<sup>nd</sup> ed. New York: McGraw Hill Book Co. p. 1-633.
- Taira H 1983. Lipid content and fatty acid composition of rice. JARQ. 16(4): 273-280.
- Tallima H, Ridi RE 2017. Arachidonic acid: physiological roles and potential health benefits
   A review. J Adv Res. 11: 33-41. Doi: 10.1016/j.jare.2017.11.004

- Tudisco R, Chiofalo B, Addi L, Presti VL, Rao R, Calabro S, Musco N, Grossi M, Cutrignelli MI, Lombardi P, Infascelli F 2015. Effect of hydrogenated palm oil dietary supplementation on milk yield and composition, fatty acids profile and stearoyl-CoA desaturase expression in goat milk. Small Rumin Res. 132: 72-78.
- Van Soest PJ, Robertson JB, Lewis BA 1991. Carbohydrate methodology, metabolism and nutritional implication in dairy cattle: methods for dietary fiber and nonstarch polysaccharides inrelation to animal. J Dairy Sci. 74:3585-3597.
- Yurchenko S, Sats A, Tatar V, Kaart T, Mootse H, Joudu I 2018. Fatty acid profile on milk from Saanen and Swedish Landrace goats. Food Chem. 254: 326-332.
- Zhang H, Ao CJ, khas-Erdene LW, Song, Zhang XF 2015. Effect of different model diets on milk composition and expression of genes related to fatty acid synthesis in the mammary gland of lactating dairy goats. J Dairy Sci. 98:1-10.