

Research Article

Fungistatic Property of *Eugenia caryophyllus* Bullock & Harrison and *Acorus calamus* Linn. Extracts Against *Candida albicans* and *Cryptococcus neoformans*

Tragoolpu K*, Thirach S*, Khamwan C**, Jatisatiern C***, Jatisatiern A***, Kunyanone N****, Punjaisee, S*

Abstract

Objective: To investigated the anti-fungal activity of two medicinal plant extracts comparing with eugenol and amphotericin B (AmB) by using the National Committee for Clinical Laboratory Standards (NCCLS) M27-P broth microdilution method.

Materials and methods: The ethanol extracts of clove (*Eugenia caryophyllus* Bullock & Harrison) and sweet flag (*Acorus calamus* Linn.), eugenol and amphotericin B were determined for minimum inhibitory concentrations (MICs) and minimum fungicidal concentrations (MFCs) against 28 clinical isolates of *Candida albicans* and 25 clinical isolates of *Cryptococcus neoformans*.

Results: The MICs of clove, sweet flag, eugenol and AmB against *C. albicans* were 17.41 ± 8.64 mg/mL, 28.8 ± 16.32 mg/mL, 12.16 ± 4.53 mg/mL and 0.23 ± 0.1 μ g/mL, respectively. The MFCs were 67.5 ± 15.39 mg/mL, >75 mg/mL, 15.4 ± 6.47 mg/mL and 0.47 ± 0.21 μ g/mL, respectively. The same extracts or antifungal drugs, which tested against *C. albicans* were also determined against *C. neoformans*. The MICs were 2.43 ± 0.95 mg/mL, 3.02 ± 1.97 mg/mL, 6.28 ± 3.4 mg/mL and 0.28 ± 0.15 μ g/mL, respectively. The MFCs were 22.22 ± 12.71 mg/mL, 30.82 ± 27.11 mg/mL, 10.06 ± 4.9 mg/mL and 0.51 ± 0.25 μ g/mL, respectively.

The results showed that *C. albicans* were statistically significant ($p < 0.01$) more susceptible to the extract of clove than sweet flag, whereas *C. neoformans* were not statistically significant susceptible to the clove extract ($p > 0.05$). Moreover, the extract of clove showed significantly ($p < 0.01$) more potent inhibitory activity against *C. neoformans* and *C. albicans* than eugenol.

Conclusion: These data indicated that the extracts of clove and sweet flag were potential fungistatic agents against yeasts whereas AmB and eugenol showed fungicidal effects. Bull Chiang Mai Assoc Sci 2001; 34: 89-97.

Key words: Clove, sweet flag, antifungal susceptibility, *Candida albicans*, *Cryptococcus neoformans*

- * Department of Clinical Microbiology, Faculty of Associated Medical Sciences, Chiang Mai university,
- ** Microbiology Section, The Central Laboratory Services Unit, Faculty of Medicine, Chiang Mai university,
- *** Department of Biology, Faculty of Science, Chiang Mai university,
- **** Microbiology section, Chiang Rai Regional Hospital

บทคัดย่อ : คุณสมบัติด้านเชื้อราของสารสกัดการพกและว่าน้ำต่อเชื้อ *Candida albicans* และ *Cryptococcus neoformans*

ขจรศักดิ์ ตระกูลพัว*, โภวิต ชิราษ*, จันทนา คำวรวณ**, ชัยวัฒน์ ชาติกาเสถียร***, อารยา ชาติกาเสถียร***, เนาวรัตน์ กันยานนท์****, สุชาติ ปันจัยสิน*

วัตถุประสงค์ : เพื่อศึกษาคุณสมบัติด้านการเจริญเติบโตของเชื้อราของสารสกัดจากพิชสมุนไพรสองชนิด เปรียบเทียบกับยูจีนอลและยาแอนไฟเทอเรชิน ปี โดยใช้วิธี Broth dilution ของ National Committee for Clinical Laboratory Standards (NCCLS) M27-P

วัสดุและวิธีการ : ทำการสกัดพิชสมุนไพรสองชนิดได้แก่ กานพกและว่าน้ำด้วยแอลกอฮอล์ แล้วนำสารสกัดทั้งสองชนิดมาทดสอบหาค่าความเข้มข้นต่ำสุดที่ยับยั้งการเจริญเติบโต (MIC) และทำลายเชื้อรา (MFC) เปรียบเทียบกับยูจีนอลและยาแอนไฟเทอเรชินโดยศึกษาคุณสมบัติดังกล่าวกับเชื้อ *Candida albicans* และ *Cryptococcus neoformans* จำนวน 28 และ 25 สายพันธุ์ ตามลำดับ

ผลการทดลอง : พบร่วมค่า MICs และ MFCs ของสารสกัดจากการพกและว่าน้ำด้วยแอลกอฮอล์ ยูจีนอล และยาแอนไฟเทอเรชิน ปี ต่อเชื้อ *C. albicans* ได้แก่ 17.41 ± 8.64 mg/mL, 28.8 ± 16.32 mg/mL, 12.16 ± 4.53 mg/mL, 0.23 ± 0.1 μ g/mL และ 67.5 ± 15.39 mg/mL, >75 mg/mL, 15.4 ± 6.47 mg/mL, 0.47 ± 0.21 μ g/mL ตามลำดับ สำหรับค่า MICs และ MFCs ของสารสกัดจากการพกและว่าน้ำด้วยแอลกอฮอล์ ยูจีนอลและยาแอนไฟเทอเรชิน ปี ต่อเชื้อ *C. neoformans* มีค่าเป็น 2.43 ± 0.95 mg/mL, 3.02 ± 1.97 mg/mL, 6.28 ± 3.4 mg/mL, 0.28 ± 0.15 μ g/mL และ 22.22 ± 12.71 mg/mL, 30.82 ± 27.11 mg/mL, 10.06 ± 4.9 mg/mL, 0.51 ± 0.25 μ g/mL ตามลำดับ

จากผลการทดลองดังกล่าวแสดงให้เห็นว่าสารสกัดจากการพกมีฤทธิ์ต้านเชื้อราต้านเชื้อ *C. albicans* (p<0.01) และไม่มีความแตกต่างกันอย่าง

มีนัยสำคัญทางสถิติสำหรับเชื้อ *C. neoformans* ($p>0.05$) นอกจากนั้นยังพบว่า สารสกัดจากการพลูมีประสิทธิภาพในการยับยั้งการเจริญเติบโตของเชื้อ *C. albicans* และ *C. neoformans* ตีกว่ายูจีนอลอย่างมีนัยสำคัญทางสถิติ ($p<0.01$) อีกด้วย

สรุปผลการทดลอง : สารสกัดจากการพลูและว่าน้ำด้วยแอลกอฮอล์มีคุณสมบัติในการยับยั้งการเจริญเติบโตของเชื้อรากีน้ำที่นำมาใช้ในการทดสอบ ขณะที่ยูจีนอลและยาแอมโพเทอเรซิน บีมีคุณสมบัติ ตีกว่าในการกำลังเชื้อราก วารสารเทคนิคการแพทย์เชียงใหม่ 2544; 34: 88-97.

คำรหัส : การพลู, ว่าน้ำ, การทดสอบความไวต่อสารต้านเชื้อราก, *Candida albicans*, *Cryptococcus neoformans*

- * ภาควิชาจุลชีววิทยาคลินิก คณะเทคโนโลยีการแพทย์ มหาวิทยาลัยเชียงใหม่,
- ** หน่วยปฏิบัติการจุลชีววิทยา งานปฏิบัติการกลางและชั้นสูตรโรค โรงพยาบาลรามาธิราณครเชียงใหม่ คณะแพทยศาสตร์ มหาวิทยาลัยเชียงใหม่,
- *** ภาควิชาชีววิทยา คณะวิทยาศาสตร์ มหาวิทยาลัยเชียงใหม่,
- **** งานจุลชีววิทยาคลินิก โรงพยาบาลศูนย์เชียงรายประจำหน่วย

Introduction

Since the incidence of fungal infections has increased significantly in the last 20 years¹. In immunocompromised patients, the emergence of candida infections with both primary drug resistance and the secondary development of azole-resistant *Candida* spp. isolates have been described^{2,3,4}. Amphotericin B has been provided for the standard treatment of the most systemic fungal infections⁵. Unfortunately, treatment with amphotericin B, especially for long-term periods, can lead to its adverse effects to patients, or development of resistant organisms during the course of therapy⁶. In the quest for new antifungal agents, low toxicity and broad spectrum fungicidal activities are needed for effective management of the infections.

Eugenia caryophyllus Bullock & Harrison

(Clove) and *Acorus calamus* Linn. (Sweet flag) have eugenol as a major constituent component. These medicinal plants have been used for the traditional medicine in Thailand and certain medical applications. Both plants have been reported to possess inhibitory properties to filamentous fungi *in vitro*^{7,8}.

Only limited knowledge is available regarding the antifungal activities of plant. Therefore, the aim of this study was to determine the antifungal activities of *Eugenia caryophyllus* Bullock & Harrison (Clove) and *Acorus calamus* Linn. (Sweet flag) extracts against clinical isolates of *C. neoformans* and *C. albicans* by broth microdilution method.

Materials and methods

Plant materials

Flowers of *Eugenia caryophyllus* Bullock

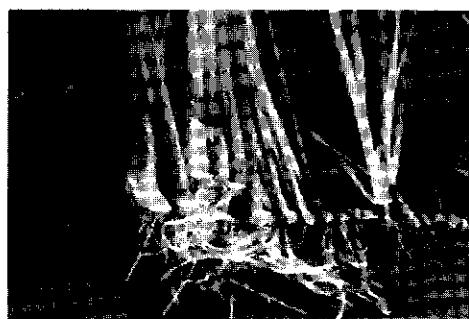
& Harrison (Clove) and rhizomes of *Acorus calamus* Linn. (Sweet flag) were selected to study (Fig.1).

Plant extracts were prepared as follows. Air dried plant materials (200 g) were finely ground. Then, the plant materials were infused in 95% ethanol and sonicated with Ultrasonic bath (Bandelin Sonorex super

RF 510H) for 30 min. The extracts were then filtered through Whatman filter paper No.1. The filtrate was evaporated and concentrated under rotary vacuum evaporator^{8,9}. The concentrated plant material was then soaked in 10 ml of 95% ethanol. Finally, the ethanolic extracts were dried, weighted and kept at -20°C in sterilized bottles.



A)



B)

Fig. 1 The medicinal plants were used in this study.

- A) Flowers of *Eugenia caryophyllus* Bullock & Harrison (Clove)
- B) Rhizomes of *Acorus calamus* Linn. (Sweet flag)

Fungal isolation

Fifty-three clinical isolates (28 of *C.albicans* and 25 of *C. neoformans*) were isolated from oral, vaginal, urine and cerebrospinal fluid of patients from Maharaj Nakorn Chiang Mai and Chiang Rai regional hospital, Thailand. The reference strain, *C.albicans* ATCC 90028, was included in all susceptibility tests as a control. The isolates were identified according to standard procedure¹⁰ and cultured on Sabouraud dextrose agar (SDA) plates (BBL, Cockeysville, Md) at 35°C for 24–48 h to ensure optimal growth before testing.

Assay medium^{11,12}

RPMI 1640 powder (BIOCHOM KG) was prepared in distilled water and adjusted pH to 7.0 with 0.165 M morpholinopropane-sulfonic acid (MOPS, Sigma). This assay medium was filter sterilized by using 0.2 µm millipore size filters (Acrodisc®32, Gelman Sciences), aliquoted and stored at 4°C until use.

Drug and medicinal plant extracts preparation

Ten serial twofold dilutions in RPMI 1640

of Amphotericin B (AmB), eugenol and two medicinal plant extracts were prepared from stock solution and arranged in rows as well as a growth control well (Without drug) and a purity control well which contained yeast-free medium¹¹. Stock solution of AmB deoxycholate (Fungizone, Squibb Industria Farmaceutica S.A.) and eugenol were prepared at 16,000 µg/mL and 500 mg/mL in dimethyl sulfoxide (Sigma), respectively. The final concentration ranges were 1.6 to 0.003 µg/mL for AmB, 50 to 0.98 mg/mL for eugenol and 75.0 to 0.15 mg/mL for both ethanol medicinal plant extracts.

Inocula preparation

Yeast inocula were prepared as previously described^{11,13,14}. Briefly, yeast was grown in SDA plate for 24 h (*C. albicans*) or 48 h (*C. neoformans*). For each isolate, five colonies were grown until the diameter of colonies was at least 1 mm. Then, the colonies were picked and suspended in 0.85% saline solution. The suspension was adjusted to the turbidity of a 0.5 McFarland standard at a wavelength of 530 nm. Quantitative colony plate count was determined on SDA to verify the inoculum size. Testing of antifungal activity was performed in 96-well round-bottom microtitration plates (NuncTM). Microdilution wells were inoculated with 100 µL of yeast suspension in RPMI 1640 medium. The final inoculum concentration of approximately 5.0×10^2 to 2.5×10^3 blastoconidia/mL for *C. albicans* and

5.0×10^3 to 2.5×10^4 blastoconidia/mL for *C. neoformans* after dilution with 100 µL of the drug solutions or extracts. The inoculated plates were incubated at 35°C, 24 h for *C. albicans* and 72 h for *C. neoformans*. Two replicated plates were used for each treatment.

Time of reading¹¹

MICs were determined after incubation for 24 and 72 h at 35°C for *C. albicans* and *C. neoformans*, respectively.

Endpoint determination

Growth of yeasts in each well was estimated visually and then scored as previously described¹³ as, 0, optically clear; 1+, slightly hazy, i.e., turbidity of more than 0–25% compared to the drug-free growth control; 2+, turbidity of more than 25 to 50% of growth control; 3+, turbidity of more than 50 to 75% of growth control; and 4+, turbidity of more than 75 to 100% of growth control.

Minimum fungicidal concentration (MFC) experiments were adapted from the method of McGinnis¹⁵. Briefly, 100 µL aliquots from tubes that showed their growth inhibition were plated on to SDA plates. The lowest drug concentration that yielded fewer yeast colonies was recorded as the MFC.

Statistical analysis

Data were analyzed and compared using student's t-test analysis.

Results

In this report, the various concentrations of ethanol crude extracts of clove, sweet flag and commercial eugenol tested were comparable to the standard AmB. The results of the experiments were summarized in Table 1 and 2. Both medicinal plant extracts have shown antifungal activities against *C. albicans* and *C. neoformans*. In Table 1, the average MICs (Mean \pm SD) of clove, sweet flag, eugenol and AmB against *C. albicans* were 17.41 ± 8.64 mg/mL, 28.8 ± 16.32 mg/mL, 12.16 ± 4.53 mg/mL and 0.23 ± 0.1 μ g/mL, respectively. Likewise, the same extracts, eugenol and antifungal drugs, which tested against *C. albicans* were also determined against *C. neoformans*. The average MICs (Mean \pm SD) were 2.43 ± 0.95

mg/mL, 3.02 ± 1.97 mg/mL, 6.28 ± 3.4 mg/mL and 0.28 ± 0.15 μ g/mL, respectively. Sweet flag showed a broad range activity against *C. albicans*, whereas eugenol showed a broad range activity against *C. neoformans*. Generally, the average MICs of eugenol, clove and sweet flag showed less effective in inhibiting the growth of *C. albicans* than *C. neoformans* (Table 1).

Moreover, the average MFCs (Mean \pm SD) of clove, sweet flag, eugenol and AmB against *C. albicans* were 67.5 ± 15.39 mg/mL, >75 mg/mL, 15.4 ± 6.47 mg/mL and 0.47 ± 0.21 μ g/mL, respectively. Similarly, the MFCs (Mean \pm SD) against *C. neoformans* were 22.22 ± 12.71 mg/mL, 30.82 ± 27.11 mg/mL, 10.06 ± 4.9 mg/mL and 0.51 ± 0.25 μ g/mL, respectively (Table 2).

Table 1 Average MICs of AmB, eugenol, clove and sweet flag extracts against *C. albicans* and *C. neoformans*.

Organisms	MICs (Mean \pm SD)			
	AmB (μ g/mL)	Eugenol (mg/mL)	Clove (mg/mL)	Sweet flag (mg/mL)
<i>C. albicans</i>	0.23 ± 0.10	12.16 ± 4.53	17.41 ± 8.64	28.80 ± 16.30
<i>C. neoformans</i> .	0.28 ± 0.15	6.28 ± 3.40	2.43 ± 0.95	3.02 ± 1.97

Table 2 Average MFCs of AmB, eugenol, clove and sweet flag extracts against *C. albicans* and *C. neoformans*.

Organisms	MICs (Mean \pm SD)			
	AmB (μ g/mL)	Eugenol (mg/mL)	Clove (mg/mL)	Sweet flag (mg/mL)
<i>C. albicans</i>	0.47 ± 0.21	15.40 ± 6.47	67.50 ± 15.39	>75.0
<i>C. neoformans</i> .	0.51 ± 0.25	10.06 ± 4.90	22.22 ± 12.71	30.82 ± 27.11

Discussion

Studies reported by Pabla *et al*⁶ and Teissedre *et al*⁷ were indicated that an essential oil was natural products extracted from plant materials, which can be used as antibacterial, antifungal, antioxidants, and anti-carcinogenic agents or to preserve and give specific flavors to foods. The susceptibility of yeast to ethanol crude extracts of clove, sweet flag was expected from eugenol, which is a mainly constituent component in both medicinal plants. The ethanol crude extracts of both plant were separated by thin layer chromatography (TLC) comparable to eugenol. The data showed that the plants and eugenol had the same Rf, 0.4. Therefore, this result indicated that eugenol was constituent in both plant. Eugenol is a derivative of phenol that effectively kills vegetative cells of bacteria and spores by causing membrane damage, leakage of cytoplasmic contents in the cells¹⁸, inhibition of enzyme activities and denaturation of protein¹⁹.

The results showed that *C. albicans* were statistically significant ($p<0.01$) more susceptible to the extract of clove than sweet flag, whereas *C. neoformans* were not statistically significant susceptible to the clove extract ($p>0.05$). Moreover, the extract of clove showed significantly ($p<0.01$) more potent inhibitory activity against *C. neoformans* than eugenol, whilst it showed significantly ($p<0.01$) less inhibitory activity against *C. albicans* than eugenol.

In conclusion, comparison of standard antimycotic agent; amphotericin B to extracts showed that extracts from both medicinal plants had low antimycotic activity. However, it is important to point out that the extracts of clove and sweet flag were potential fungistatic agents against yeasts whereas AmB and eugenol showed fungicidal activities.

Acknowledgment

We thank Assoc. Prof. Rujapa Nimsung of the Associated Medical Sciences faculty, Chiang Mai University for collaborating with us in the freeze-dried medicinal plant preparation and Dr. Yingmanee Tragooolpua for her assistance in the preparation of this manuscript.

References

1. Poeta MD, Bixel AS, Barchiesi F, *et al*. In vitro activity of dicationic aromatic compounds and fluconazole against *Cryptococcus neoformans* and *Candida* spp. *J Antimicrob Chemother* 1999; 44: 223-8.
2. Cameron ML, Schell WA, Brunch S, *et al*. Correlation of *in vitro* fluconazole resistance of *Candida* isolates in relation to therapy and symptoms of individuals seropositive for human immunodeficiency virus type 1. *Antimicrob Agents Chemother* 1993; 3; 2449-53.
3. Pfaller MA, Rhine-Chalberg AJ, Redding SW, *et al*. Variations in fluconazole susceptibility and electrophoretic karyotype

among oral isolates of *Candida albicans* from AIDS and oral candidiasis. *J Clin Microbiol* 1994; 32; 59-64.

4. Willocks L, Leen CLS, Brette RP, et al. Fluconazole resistance in AIDS patients. *J Antimicrob Chemother* 1991; 28; 937-9.
5. Medoff G, Kobayashi GS. Strategies in the treatment of systemic fungal infections. *N Engl J Med* 1980; 302;145-55.
6. Kovacicova G, Hanzen J, Pisarcikova M, et al. Nosocomial fungemia due to amphotericin B-resistant *Candida* spp. in three pediatric patients after previous neurosurgery for brain tumors. *J Infect Chemother* 2001; 7; 45-8.
7. Hitokoto H, Morozumi S, Wauke T, Sakai S, Kurata H. Inhibitory effect of spices on growth and toxin production of toxigenic fungi. *Appl Environ Microb* 1980; 39; 818-22.
8. Tragoopua K. Effect of the extract from eight species of medicinal plants on growth of selected plant pathogenic molds and dermatophytes. M.Sc. Thesis, Chiang Mai University 1996.
9. Fabry W, Okemo P, Ansorg R. Fungistatic and fungicidal activity of East African medicinal plants. *Mycoses* 1996; 39; 67-70.
10. Mahon CR, Manuselis G,Jr. *Textbook of diagnostic microbiology*. Philadelphia: WB Saunders, 1995.
11. Cormican MG, Pfaffer MA. Standardize of antifungal susceptibility testing. *J Antimicrob Chemother* 1996; 38; 561-78.
12. National Committee for Clinical Laboratory Standards. Reference method for broth dilution antifungal susceptibility testing of yeasts. Approved standard M27-A. Wayne, PA. , National Committee for Clinical Laboratory Standards, 1997.
13. Anaissie EJ, Paetznick VL, Ensign LG, et al. Microdilution Antifungal susceptibility testing of *Candida albicans* and *Cryptococcus neoformans* with and without agitation: an eight-center collaborative study. *Antimicrob Agents Chemother* 1996; 40; 2387-91.
14. Archiesi F, Colombo AL, McGough DA, Ronald MD Comparative study of macrodilution and microdilution techniques for *in vitro* antifungal susceptibility testing of yeasts by using the National Committee for Clinical Laboratory Standards proposed standard. *J Clin Microbiol* 1994; 32; 2494-500.
15. McGinnis MR. Susceptibility Testing and Bioassay Procedure. In : *Laboratory Handbook of Medical Mycology*. New York : Academic Press, 1980: 431.
16. Pabla T, Gulati MS, Mohan U. Evaluation of antimicrobial efficacy of various root canal filling materials for primary teeth. *J Indian Soc Pedod Prev Dent* 1997; 15; 134-40.
17. Teissedre PL, Waterhouse AL. Inhibition of oxidation of human low-density lipoproteins by phenolic substances in

different essential oils varieties. *J Agric Food Chem* 2000; 48; 3801–5.

18. Lim DV. *Microbiology*. 2nd edition. Boston: WCB McGraw–Hill, 1998.

19. Sooksri-ngam B. *Microbial inhibition from some spices*. M.Sc. Thesis Kasetsart University, 1985.