

Identification of Specific Biomarker Genes for Separating Intrahepatic Cholangiocarcinoma Type

Thitiluck Swangsri¹
Thanyaphorn Junkree¹
Sopit Wongkham^{2,3}
Panee Chaksangchaichot¹
Songsak Petmitr¹

Abstract Intrahepatic cholangiocarcinoma (ICC) is one of the most lethal liver cancers. ICC is classified into 3 types: mass-forming (MF), periductal infiltrating (PI), and intraductal growth (IG). Among these, the PI and IG types are located in the bile duct, so that histopathologic diagnosis may be quite difficult. This study aims to identify the specific biomarkers for distinguish between PI and IG types. The top five up-regulated genes from PI (C19ORF33, UPK1B, CTHRC1, GREM1, KLK11) and IG type (CFTR, SLC5A1, MUC17, BDKRB2, CCL18) were examined in ICC samples (11 PI and 11 IG samples) using SYBR green-based real-time reverse transcription-PCR (qRT-PCR). Among these genes, the results showed that the CTHRC1 gene of the PI type identified with the highest up-regulation in PI (72.7%) and 27.3% in IG samples ($P=0.034$), whereas the CFTR gene of the IG type showed the highest up-regulation in 72.7% of IG and 9.1% of PI samples ($P=0.001$). Hence, the up-regulation of CTHRC1 and CFTR may serve as biomarkers for separating PI from IG types, respectively, and is also a potential diagnostic in ICC. (*Thai Cancer J* 2020;40:76-85)

Keywords: intrahepatic cholangiocarcinoma, biomarker, genes

¹Department of Molecular Tropical Medicine and Genetics, Faculty of Tropical Medicine, Mahidol University

²Department of Biochemistry, Faculty of Medicine

³Cholangiocarcinoma Research Institute, Khon Kaen University

For correspondence: thitiluck.swa@mahidol.ac.th

การศึกษาตัวบ่งชี้ทางชีวภาพในการจำแนกชนิดของมะเร็งท่อน้ำดีภายในตับ

โดย ฐิติลักษณ์ สว่างศรี¹, ชัยพร จันทร์¹, โสพิศ วงศ์คำ^{2,3}, พาณี จักรแสงชัยโชติ¹, ทรงศักดิ์ เพ็ชรมิตร¹

¹ภาควิชาชีวโมเลกุลและพันธุศาสตร์โรคเขตร้อน คณะเวชศาสตร์เขตร้อน มหาวิทยาลัยมหิดล, ²ภาควิชาชีวเคมี คณะแพทยศาสตร์, ³สถาบันวิจัยมะเร็งท่อน้ำดี มหาวิทยาลัยขอนแก่น

บทคัดย่อ มะเร็งท่อน้ำดีภายในตับเป็นมะเร็งชนิดที่ร้ายแรงที่สุดชนิดหนึ่ง แบ่งได้เป็น 3 ลักษณะตามพยาธิสภาพ ได้แก่ massforming (MF), periductal infiltrating (PI) และ intraductal growth (IG) และเนื่องจากมะเร็งท่อน้ำดีภายในตับชนิด PI และ IG เกิดบริเวณภายในท่อน้ำดีจึงอาจทำให้ตรวจหาด้วยวิธีทางจุลพยาธิวิทยาได้ค่อนข้างยาก ดังนั้นการศึกษานี้จึงมีวัตถุประสงค์เพื่อหาตัวบ่งชี้ทางชีวภาพในการจำแนกมะเร็งท่อน้ำดีภายในตับ โดยคัดเลือกยีนที่มีการแสดงออกสูงสุด 5 ลำดับแรกจากแต่ละชนิด ได้แก่ กลุ่ม PI: C19ORF33, UPK1B, CTHRC1, GREM1, KLK11 กลุ่ม IG: CFTR, SLC5A1, MUC17, BDKRB2, CCL18 มาทดสอบกับตัวอย่างมะเร็งท่อน้ำดีภายในตับ (ชนิด PI 11 ตัวอย่างและชนิด IG 11 ตัวอย่าง) ด้วยเทคนิคเรียลไทม์อาร์ทีพีซีอาร์ (SYBR green-based real-time reverse transcription-PCR) ผลการศึกษาพบว่ายีน CTHRC1 จากกลุ่ม PI มีการแสดงออกสูง (up-regulated) มากที่สุด (ร้อยละ 72.7) ในตัวอย่างชนิด PI และพบร้อยละ 27.3 ในตัวอย่างชนิด IG ($P=0.034$) ในขณะที่ยีน CFTR จากกลุ่ม IG มีการแสดงออกสูงมากที่สุด (ร้อยละ 72.7) ในตัวอย่างชนิด IG ส่วนในตัวอย่างชนิด PI พบเพียงร้อยละ 9.1 ($P=0.001$) ดังนั้นยีน CTHRC1 จากกลุ่ม PI และยีน CFTR จากกลุ่ม IG สามารถนำมาใช้เป็นตัวแทนของตัวบ่งชี้ทางชีวภาพในการจำแนกความแตกต่างระหว่างมะเร็งท่อน้ำดีภายในตับชนิด PI และ IG ซึ่งผลที่ได้นี้สามารถนำมาใช้ต่อยอดในการวินิจฉัยมะเร็งท่อน้ำดีภายในตับได้อย่างมีประสิทธิภาพต่อไปในอนาคต (วารสารโรคมะเร็ง 2563;40:76-85)

คำสำคัญ: มะเร็งท่อน้ำดีภายในตับ ตัวบ่งชี้ทางชีวภาพ ยีน

Introduction

ICC is the second most common primary hepatobiliary cancer globally and is a major cause of cancer-related deaths that shows no indication of a decrease in mortality rate¹⁻³. In developing countries especially in Thailand has the highest incidence of ICC in the world, which may be related to *Opisthorchis viverrini* infection⁴⁻⁶. The prevalence of liver fluke infection in northeast Thailand is about 317.6 per 100,000 person-years⁶.

In detection, it should be emphasized that imaging is not always reliable for the diagnosis of some ICC. For instance, ICC <2 cm in size mimic HCC because of the absence of the progressive enhancement pattern⁷. Moreover, it may quite difficult to use only histopathologic diagnostic for differential three types of ICC (MF, PI and IG) which are located in bile duct. From the above problems, there are several studies searching for novel biomarkers that are likely to be helpful in diagnosis,

characterization, therapy selection including separating type. A deep understanding of the relative relevance of each biomarker will be a key to successfully diagnose diseases, therapy and monitoring⁸⁻¹¹.

To date, 2821 up-regulated genes in ICC for Thai patients have been detected. Moreover, 531 and 1535 genes were specific up-regulated in PI and IG types, respectively^{12,13}. The present study aims to validate these selected specific genes especially in PI and IG types, which may be used as a molecular biomarker to identify type of ICC.

Materials and Methods

Patients and clinical features

ICC fresh tumor samples (11 PI and 11 IG samples) including with corresponding normal samples were leftover specimen collection at Department of Molecular Tropical Medicine and Genetics, Faculty of Tropical Medicine, Mahidol University, Thailand which transferred from Cholangiocarcinoma Research Center, Faculty of Medicine, Khon Kaen University, Thailand. The study was approved by the Ethics Committee of the Faculty of Tropical Medicine, Mahidol University, Thailand (MUTM 2008-004-02) and (MUTM 2014-057-03).

Sample processing

Total RNA was isolated by TRIzol[®] reagent (Invitrogen, USA) according to the manufacturer's instructions. The total amount of RNA isolated was quantified using a Nanodrop 2000 c spectrophotometer (Thermo Fisher Scientific, USA) OD₂₆₀ measurements, and its quality evaluated by visualizing specific bands (18S and 28S rRNA) using 1.5% gel electrophoresis. First strand cDNA

was performed using Superscript[®] VILO™ cDNA Synthesis kits (Invitrogen, USA) and purified by DNAClear™ cDNA Purification Kit (Applied Biosystems, USA). The quantity of purified cDNA was measured by Nanodrop2000 c spectrophotometer and qualified using conventional PCR with GAPDH primer.

Validation of candidate gene expression by SYBR green-based real-time reverse transcription-PCR (qRT-PCR)

Primers were designed by Primer-BLAST program (NCBI) using nucleotide sequences from the NCBI database. The nucleotide sequences of all primers are shown in Table 1. The LightCycler[®] FastStart DNA Master SYBR Green I (Roach, Germany) was used to perform SYBR green qPCR. Each 10 µl of reaction mixture contained 5 µl of 2X MasterMix SYBR Green, 0.3 µM of forward primer, 0.3 µM of reverse primer, 3.4 µl of sterile distilled water and 1 µl of cDNA. The reactions were carried out using a Roche Lightcycler[®] 480 Real-Time PCR System (Roche, Germany), with the following cycling conditions: pre-incubation at 95°C for 5 minutes, followed by 45 cycles of denaturation at 95°C for 10 seconds, annealing temperature (specific to the primer) for 20 seconds, and extension at 72°C for 20 second. The $2^{-\Delta\Delta CT}$ method was used to calculate relative quantification in gene expression. Over-expression of mRNA was specified as N-fold change ≥ 1.5 , normal mRNA expression was specified as N-fold change (range 0.5001-1.4999), and under-expression was specified as N-fold change ≤ 0.5 . All samples were performed at least twice^{20,21}.

Table 1 The oligonucleotide sequences of PI and IG types

Type	Gene	Sequence (5' to 3')
PI	C19ORF33-F	AGCATTCCCTCCAACGGGCA
	C19ORF33-R	GTCGAACTCCATGGCGGTAA
	UPK1B-F	AGCCTCTACCCACTGCTTGA
	UPK1B-R	GGAAGAGGTTGGGTGTGAAA
	CTHRC1-F	TTGTTCAAGTGGCTCACTTCG
	CTHRC1-R	TTCAATGGGAAGAGGTCCTG
	GREM1-F	GCTCTGGCATTGAGAGAACC
	GREM1-R	AAATTCGCCTAGCGTGAGAA
	KLK11-F	ATCACCATCATTGAGCACCA
	KLK11-R	CCCAGGAGATAATGCCTTGA
IG	CFTR-F	ACAGGATGGTTCCTTGATG
	CFTR-R	GGGCTGTCCTGTGACAATTT
	SLC5A1-F	CAACATCGCCTATCCAACCT
	SLC5A1-R	TAAACAACCTTCCGGCAATC
	MUC17-F	CGCTTTCTGCAACCAGTACA
	MUC17-R	ACTGGAGTGAGCAGGAGGAA
	BDKRB2-F	GGTTGTGCTGCTGCTATTCA
	BDKRB2-R	GCACACTCCCTGGTACACCT
	CCL18-F	AGAAGGAGGCCAGGAGTTGT
	CCL18-R	GTGGAATCTGCCAGGAGGTA

F: forward primer, R: reverse primer

Statistical analysis

The association between gene expression of PI and IG types were performed with SPSS statistical software. Differences in expression levels in these types were analyzed using the student t-test, and $P < 0.05$ was considered statistically significant.

Results

Among top five of up-regulated genes selected from each PI and IG types, CTHRC1 gene was found as the most up-regulated gene in PI sample (72.7%) followed by C19ORF33 (63.6%), GREM1 (54.5%), KLK11 (45.5%), MUC17 (45.5%), UPK1B (36.4%), BDKRB2 (27.3%), CCL18 (27.3%), SLC5A1 (18.2%), and CFTR (9.1%), as shown in Figure 1.

Table 2 Gene expression patterns of CTHRC1 and CFTR
in PI and IG samples

sample	No.	Gene expression patterns	
		CTHRC1	CFTR
PI	1	+	-
	2	+	-
	3	-	-
	4	+	+
	5	-	-
	6	+	-
	7	+	-
	8	-	-
	9	+	-
	10	+	-
	11	+	-
IG	1	-	-
	2	-	-
	3	-	+
	4	-	+
	5	+	+
	6	-	-
	7	+	+
	8	-	+
	9	-	+
	10	-	+
	11	+	+

+ = up-regulated, - = not up-regulated

Meanwhile, the result of up regulated gene in IG sample show that CFTR gene is the most predominant gene which was found in 8 from 11 samples (72.7%) followed by MUC17 (63.6%), BDKRB2 (63.6%), GREM1 (63.6%), C19ORF33 (54.5%), UPK1B (54.5%), CCL18 (36.4%), KLK11 (36.4%), SLC5A1 (27.3%), and CTHRC1 (27.3%), as shown in Figure 2.

Of 22 ICC samples (11 PI and 11 IG samples), the up-regulated CTHRC1 gene was found in 8/11 (72.7%) of PI samples but was

detected at relatively low frequency in 3/11 (27.3%) of IG samples ($P=0.034$), whereas up-regulate of CFTR gene was presented only in 1 (9.1%) of PI sample and were identified with high frequency in 8/11 (72.7%) of IG samples ($P=0.001$), the data are shown in Table 2 and Figure 3.

Discussion and Conclusion

ICC is the cancer of the intrahepatic bile ducts and together with hepatocellular carcinoma (HCC), which is the majority of primary liver cancers¹⁻³. Previous studies have raised some concern about the incidence and mortality related to ICC, whereas the incidence of extrahepatic cholangiocarcinoma (ECC) has remained relatively stable over time¹⁴⁻¹⁶. Patients with early stage of ICC are usually asymptomatic. Moreover, the clinical presentation is non-specific and difficult to establish diagnosis¹⁷. Therefore, the identification of ICC novel biomarkers are the keys of successful early diagnosis and it is also useful for effective treatment of patients with CCA.

In the present study, of top five of up-regulated genes from either PI or IG type of ICC determined by a previous report¹² were validated by SYBR green qRT-PCR showed that CTHRC1 and CFTR genes could be the representative of biomarkers for identification PI and IG types, respectively. CTHRC1 was the predominant gene from PI type, which showed significantly different of up-regulated gene between PI and IG samples. Previous studies revealed that collagen triple helix repeat containing 1 is a protein that in humans is encoded by the CTHRC1 gene and it has been

identified as a cancer-related gene¹⁸. Numerous reports showed the association between expression of CTHRC1 and cancer proliferation, invasion, metastasis, clinical stages, tumor size, including nodal metastasis. Moreover, up-regulation of CTHRC1 is a positive indicator of poor

prognosis in malignant tumor patients including lung cancer, epithelial ovarian cancer, colorectal cancer, hepatocellular carcinoma and breast cancer¹⁹⁻²³.

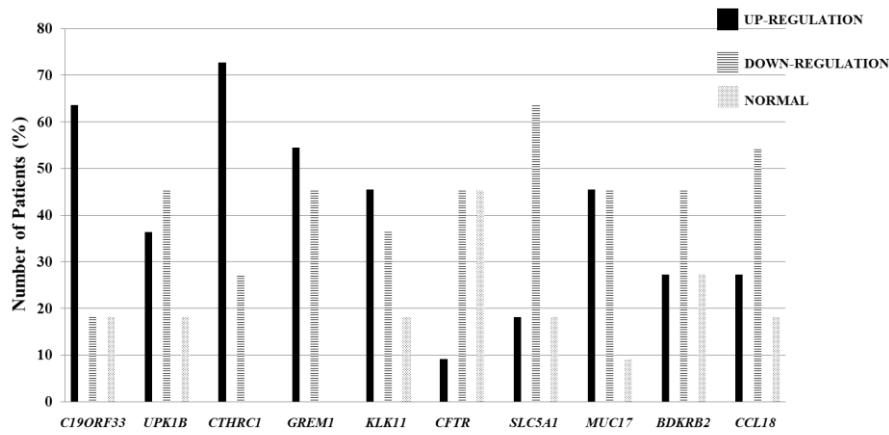


Figure 1 Differential gene expression in PI samples by SYBR Green qRT-PCR

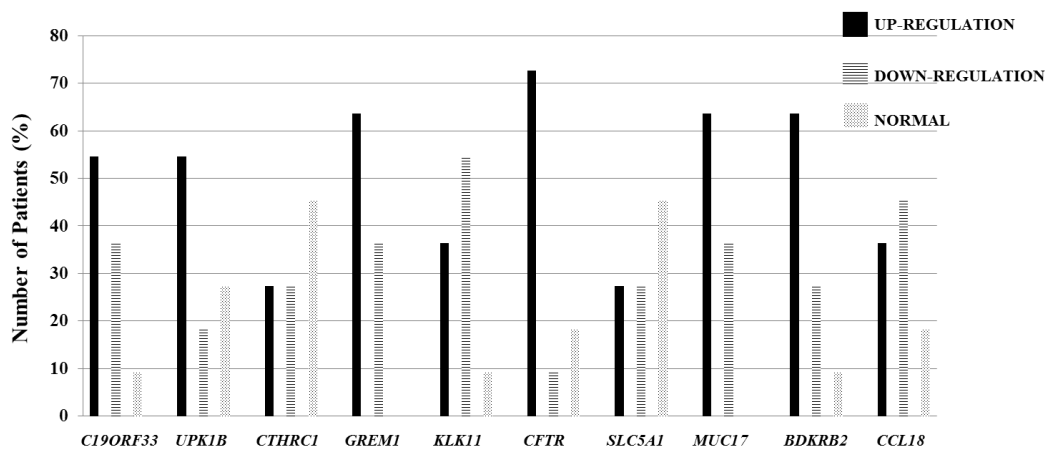


Figure 2 Differential gene expression in IG samples by SYBR Green qRT-PCR

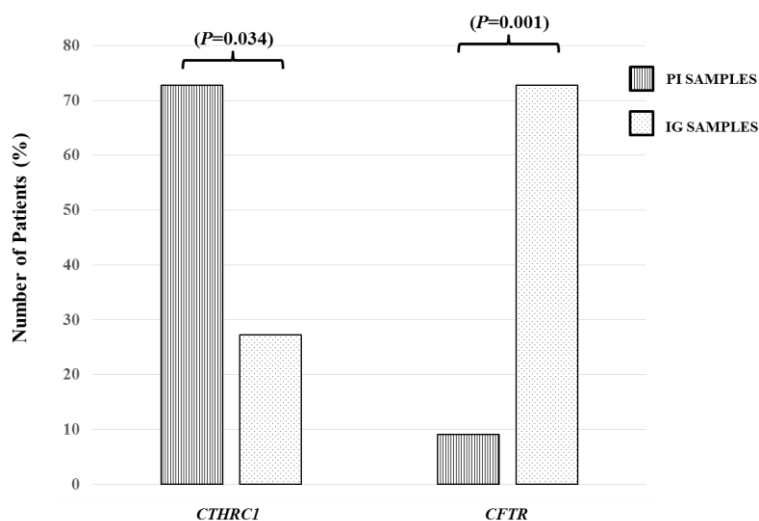


Figure 3 Comparison between up-regulation of CTHRC1 and CFTR genes in PI and IG samples by SYBR

Green qRT-PCR

A series of clinical studies have demonstrated that CTHRC1 protein and mRNA expression were detected ubiquitously in many human solid tumors²⁴, and the overexpression of CTHRC1 in tumor cells increased the characteristics of malignant aggressiveness including increased tumor growth, invasion and metastasis. Furthermore, our results provided experimental evidence that up-regulation of CTHRC1 gene could be found in ICC. Therefore, CTHRC1 gene may be developed as a potential crucial biomarker for detection of PI type in the ICC cases in Thai patients.

The CFTR gene was the most dominant gene from IG type and the up-regulation of CFTR gene was significantly different between IG and PI samples. Previous studies have suggested that cystic fibrosis transmembrane conductance regulator (CFTR) is a membrane protein and chloride channel in vertebrates that is encoded by the CFTR gene, which it may be associated with several cancers^{25,26}. It has been shown the evidence of CFTR gene expression could serve as a biomarker for lung cancer metastasis and prognosis, a prognostic predictor in breast cancer and cervical cancer²⁷⁻²⁹.

Previous research suggested that CFTR may have a more generic function in development of cancer, both in vitro and in vivo studies²⁸. Additionally, based on previous observation showed the CFTR gene provides instructions for making a protein called the CFTR protein, which is a membrane of glycoprotein expressed in certain epithelia, liver and predominantly found in the intrahepatic bile duct cells, where it plays a role in ductular bile formation^{25,30}. This information is consistent to other study which reported that CFTR is expressed in the epithelia of the intrahepatic and

extrahepatic bile ducts and the gallbladder and the CFTR protein is localized to the apical domain of these cells²⁵. In particular, the report³¹ revealed that CFTR expression was enhanced in patients with hepatolithiasis may support the supposition that CFTR is causally related to intrahepatic stones formation³². Interestingly, it is widely known that longstanding intrahepatic stones lead to ICC³³. Moreover, this study indicated that the CFTR gene could be the potential biomarker for identification IG type of ICC.

From our knowledge, several studies have been searching for a novel biomarker for detection or separating type of cancer using immunohistochemical assay (IHC), fluorescence in situ hybridization (FISH) technique, including qRT-PCR. Interestingly, the previous studies showed that qRT-PCR has the potential to become standard in terms of its performance, accuracy, sensitivity, broad dynamic range, and high throughput capacity³⁴. Moreover, qRT-PCR is more convenient, easier, and rapid compared to IHC and FISH^{35,36}.

In summary, our observations indicated that the up-regulation of CTHRC1 and CFTR genes were mainly exhibited in PI and IG types of ICC and these genes may be developed as valuable prognosis biomarkers for PI and IG types, respectively. However, a greater number of cases should be included to make the prospective studies valid in the future. Interestingly, to the best of our knowledge, our data have provided clinical findings for the first time that CTHRC1 and CFTR genes expression are well correlated with ICC.

Acknowledgement

This study was supported by Mahidol University; the Dean's Research Fund, Faculty of Tropical Medicine, Mahidol University.

References

1. Khan SA, Thomas HC, Davidson BR, Taylor-Robinson SD. Cholangiocarcinoma. *Lancet* 2005;366:1303-14.
2. Ustundag Y, Bayraktar Y. Cholangiocarcinoma: a compact review of the literature. *World J Gastroenterol* 2008;14:6458-66.
3. Mosconi S, Beretta GD, Labianca R, Zampino MG, Gatta G, Heinemann V. Cholangiocarcinoma. *Crit Rev Oncol Hematol* 2009;69:259-70.
4. Thamavit W, Pairojkul C, Tiwawech D, Shirai T, Ito N. Strong promoting effect of *Opisthorchis viverrini* infection on dimethyl nitrosamine-initiated hamster liver. *Cancer Lett* 1994;78:121-5.
5. Sripa B, Pairojkul C. Cholangiocarcinoma: lessons from Thailand. *Curr Opin Gastroenterol* 2008;24:349-56.
6. Sriamporn S, Pisani P, Pipitgool V, Suwanrungruang K, Kamsa-ard S, Parkin DM. Prevalence of *Opisthorchis viverrini* infection and incidence of Cholangiocarcinoma in Khon Kaen, Northeast Thailand. *Trop Med Int Health* 2004;9:588-94.
7. Rimola J, Forner A, Reig M, Vilana R, de Lope CR, Ayuso C, et al. Cholangiocarcinoma in cirrhosis: absence of contrast washout in delayed phases by magnetic resonance imaging avoids misdiagnosis of hepatocellular carcinoma. *Hepatology* 2009;50:791-8.
8. Hansel DE, Rahman A, Hidalgo M, Thuluvath PJ, Lillemoe KD, Shulic KR, et al. Identification of novel cellular targets in biliary tract cancers using global gene expression technology. *Am J Pathol* 2003; 163:217-29.
9. Jinawath N, Chamgramol V, Furukawa Y, Obama K, Tsunoda T, Sripa B, et al. Comparison of gene expression profiles between *Opisthorchis viverrini* and Non-*Opisthorchis viverrini* associated human intrahepatic cholangiocarcinoma. *Hepatology* 2006;44:1025-38.
10. Obama K, Ura K, Li M, Katagiri T, Tsunoda T, Nomura A, et al. Genome-wide analysis of gene expression in human intrahepatic cholangiocarcinoma. *Hepatology* 2005;41: 1339-48.
11. Wan AG, Yoon SY, Oh Jh, Jeon YJ, Kim M, Kim JM, et al. Identification of intrahepatic cholangiocarcinoma related genes by comparison with normal liver sample using expressed sequence tags. *Biochem Biophys Res Commun* 2006;345:1022-32.
12. Subrungruang I, Thawornkuno C, Petmitr CP, Pairojkul C, Wongkham S, Petmitr S. gene expression profiling of intrahepatic Cholangiocarcinoma. *Asian Pac J cancer Prev* 2013;14:557-63.
13. Subrungruang I. Study of genetic profile in cholangiocarcinoma [Ph.D. (Tropical Medicine) Thesis in Tropical Nutrition and Food Science]. Bangkok: Faculty of Graduate Studies, Mahidol University; 2012.
14. Patel T. Increasing incidence and mortality of primary intrahepatic cholangiocarcinoma in the United States. *Hepatology* 2001;33:1353-7.
15. Shaib YH, Davila JA, McGlynn K, El-Serag HB. Rising incidence of intrahepatic cholangiocarcinoma in the United States: a true increase? *J Hepatol* 2004;40:472-7.

16. Welzel TM, McGlynn KA, Hsing AW, O'Brien TR, Pfeiffer RM. Impact of classification of hilar cholangiocarcinomas (Klatskin tumors) on the incidence of intra- and extrahepatic cholangiocarcinoma in the United States. *J Natl Cancer Inst* 2006;98:873-5.
17. Bridgewater J, Galle PR, Khan SA, Llovet JM, Park JW, Patel T, et al. Guidelines for the diagnosis and management of intrahepatic cholangiocarcinoma. *J Hepatol* 2014;60:1268-89.
18. Jiang N, Cui Y, Liu J, Zhu X, Wu H, Yang Z, et al. Multidimensional roles of collagen triple helix repeat containing 1 (CTHRC1) in malignant cancers. *J Cancer* 2016 ;7: 2213-20.
19. Ke Z, He W, Lai Y, Guo X, Chen S, Li S, et al. Overexpression of collagen triple helix repeat containing 1 (CTHRC1) is associated with tumor aggressiveness and poor prognosis in human non-small cell lung cancer. *Oncotarget* 2014; 5: 9410-24.
20. Hou M, Cheng Z, Shen H, He S, Li Y, Pan Y, et al. High expression of CTHRC1 promotes EMT of epithelial ovarian cancer (EOC) and is associated with poor prognosis. *Oncotarget* 2015;6:35813-29.
21. Kim HC, Kim YS, Oh HW, Kim K, Oh SS, Kim JT, et al. Collagen triple helix repeat containing1 (CTHRC1) acts via ERK-dependent induction of MMP9 to promote invasion of colorectal cancer cells. *Oncotarget* 2014;5:519-29.
22. Chen YL, Wang TH, Hsu HC, Yuan RH, Jeng YM. Overexpression of CTHRC1 in hepatocellular carcinoma promotes tumor invasion and predicts poor prognosis. *PLoS one* 2013;8:1-10.
23. Kim JH, Baek TH, Yim HS, Kim KH, Jeong SH, Kang HB, et al. Collagen triple helix repeat containing-1 (CTHRC1) expression in invasive ductal carcinoma of the breast: the impact on prognosis and correlation to clinic opathologic features. *Pathol Oncol Res* 2013;19:731-7.
24. Tang L, Dai DL, Su M, Martinka M, Li G, Zhou Y. Aberrant expression of collagen triple helix repeat containing 1 in human solid cancers. *Clin Cancer Res* 2006;12: 3716-22.
25. Cohn JA, Strong TV, Picciotto MR, Naim AC, Collins FS, Fitz JG. Localization of the cystic fibrosis transmembrane conductance regulator in human bile duct epithelial cells. *Gastroenterology* 1993; 105:1857-64.
26. Warren N, Holmes JA, al-Jader L, West RR, Lewis DC, Padua RA. Frequency of carriers of cystic fibrosis gene among patients with myeloid malignancy and melanoma. *BMJ* 1991;302:760-1.
27. Padua RA, Warren N, Grimshaw D, SmithM, Lewis C, Whittaker J, et al. The cystic fibrosis delta F508 gene mutation and cancer. *Hum Mutat* 1997;10:45-8
28. Li J, Zhang JT, Jiang X, Shi X, Shen J, Feng F, et al. The cystic fibrosis transmembrane conductance regulator as a bio marker in non-small cell lung cancer. *Int J Oncol* 2015;46:2107-15.
29. Zhang JT, Jiang XH, Xie C, Cheng H, Da Dong J, Wang Y, et al. Downregulation of CFTR promotes epithelial-to-mesenchymal transition and is associated with poor prognosis of breast cancer. *Biochim Biophys Acta* 2013;1833:2961-9.

30. Peng X, Wu Z, Yu L, Li J, Xu W, Chan HC, et al. Overexpression of cystic fibrosis transmembrane conductance regulator (CFTR) is associated with human cervical cancer malignancy, progression and prognosis. *Gynecol Oncol* 2012;125:470-6.
31. Pall H, Zielenski J, Jonas MM, DaSilva DA, Potvin KM, Yuan XW, et al. Primary sclerosing cholangitis in childhood is associated with abnormalities in cystic fibrosis-mediated chloride channel function. *J Pediatr* 2007;151:255-9.
32. Shoda J, Kano M, Asano T, Irimura T, Ueda T, Iwasaki R, et al. Secretory low-molecular weight Phospholipases A2 and their specific receptor in bile ducts of patients with intrahepatic calculi: Factors of chronic proliferative cholangitis. *Hepatology* 1999;29:1025-36.
33. Kim HJ, Lee SK, Kim MH, Son JM, Lee SS, Park JS, et al. Cystic fibrosis transmembrane conductance regulators (CFTR) in biliary epithelium of patients with hepatolithiasis. *Dig Dis Sci* 2002;47:1758-65.
34. Mori T, Sugiyama M, Atomi Y. Gallstone disease: Management of intrahepatic stones. *Best Pract Res Clin Gastroenterol* 2006;20:1117-37.
35. Ntoulia M, Kaklamanis L, Valavanis C, Kafousi M, Stathopoulos E, Arapantoni P, et al. HER-2 DNA quantification of paraffin-embedded breast carcinomas with Light Cycler real-time PCR in comparison to immunohistochemistry and chromogenic in situ hybridization. *Clin Biochem* 2006;39:942-6.
36. Ghani N, Westenberg D. Quantitative real-time PCR assay with immunohistochemical evaluation of HER2/neu oncogene in breast cancer patients and its correlation with clinicopathological findings. *Indian J Pathol Microbiol* 2020;63:S123-8.
37. Nistor A, Watson PH, Pettigrew N, Tabiti K, Dawson A, Myal Y. Real-time PCR complements immunohistochemistry in the determination of HER-2/ neu status in breast cancer. *BMC Clin Pathol* 2006;6:1-8.