

# Stonebreaker activity review of seed-under-leaf, *Phyllanthus niruri*

Wanee Jiraungkoorskul, PhD.<sup>1\*</sup> 

<sup>1</sup> Pathology Division, Department of Preclinical Sciences, Faculty of Medicine,  
BangkokThonburi University, Bangkok, Thailand 10170

***Phyllanthus niruri*** (Family: Phyllanthaceae) known as a seed-under-leaf, is one of the traditional medicinal plants. It is widely used in anti-nephrolithiasis and anti-urolithiasis or stone removal treatment since it is able to modify the structure, shape, and texture of the stone to a smoother and probably more fragile form, which could contribute to the elimination and dissolution of stone. Further, it has been shown to possess anticancer, antidiabetic, anti-hepatotoxic, anti-hypertensive, anti-inflammatory, antimicrobial, and antioxidant properties. Its phytochemical substances are alkaloids, coumarins, flavonoids, saponins, tannins, and terpenoids. The other important constituents are astragalin, geranin, hypophyllanthin, nirtetralin, phyllanthin, phyllochrysin, phylltetralin, and quercetin, and all of these contribute to its remedial properties. This is a brief overview of the stonebreaker activity of *P. niruri* to further provide an up-to-date review showing its importance.


**Keywords:** Herb, *Phyllanthus niruri*, Plant, Seed-under-leaf, Stonebreaker

**Received:** 20 Feb 2022 | **Revised:** 15 Mar 2022 | **Accepted:** 1 April 2022

**J Med Glob 2022 May; 1(1)**

**Website:** <https://he01.tci-thaijo.org/index.php/JMedGlob/>

**\* Correspondence to:**

Wanee Jiraungkoorskul,  
Pathology Division, Department of Preclinical Sciences,  
Faculty of Medicine, BangkokThonburi University,  
Bangkok, Thailand 10170  
E-mail: [wannee.jir@bkkthon.ac.th](mailto:wannee.jir@bkkthon.ac.th)  
 ORCID: 0000-0002-3070-280X

## INTRODUCTION

The prevalence and incidence of stones, including nephrolithiasis and urolithiasis, are reported to be increasing globally. Romero et al. (2010) [1] reported these global increases are seen across age, race, sex, diet, and climate factors. National Health and Nutrition Examination Survey reported the prevalence of renal stones in US adults rose significantly from 5.2% in 1994 to 8.8% in 2012. Moreover, among men, the prevalence of stones was 10.6% compared with 7.1% among women [2]. Patients with renal stones often have a benign course, but life-threatening complications like acute

renal injury and infection can arise [3]. Moreover, the financial burden from medical expenditures and lost productivity is [4]. After accounting for increases in population and stone prevalence, obesity, diabetes, and population rates will contribute to an estimated \$1.24 billion per year increase in the cost of renal stones by 2030 [5]. Even the advanced method and technology for stone removal or the treatment of stone is available, like open surgery, nephrostomy [6], and shock wave lithotripsy [7], it has their own limitation involving highly invasive surgeries, a long in-hospital recovery period, several side effects as even periodical reoccurrence of stones in a few. The alternative system of medicine usually employs natural sources or green medicine or medicinal plants due to their safety, efficacy, cultural acceptability, and lesser side effects as compared to synthetic drugs [8,9]. The plant that find very useful in the treatment of stone for the example *Abutilon indicum* [10], *Ageratum conyzoides* [11], *Borhaavia diffusa* [12], *Capparis decidua* [13], *Cassia tora* [14], *Launaea*

**How to cite this article:** Jiraungkoorskul. W. Stonebreaker activity review of seed-under-leaf, *Phyllanthus niruri*. J Med Glob 2022; 1(1): 59-65.



**Figure 1.** *Phyllanthus niruri* morphology (A) leaf, stem and root; (B) flower; (C-D) seed.

*procumbens* [15], *Morus alba* [16], *Phyllanthus niruri* [17], *Solanum xanthocarpum* [18].

## Phyllanthus niruri

### Plant description

*P. niruri* is an erect annual herb, growing 40-70 cm in height and having to ascend herbaceous branching, it is quite glabrous and branching at the base (Figure 1). **Stem:** horizontal branches and height of 30-60 cm, 1-2.5 mm width. **Leaves:** numerous, small, green, subsessile, closely arranged, elliptic oblong-shaped, obtuse, having short petiole and stipules present, they are arranged alternatively on each side of the stem. **Flowers:** yellowish, small, numerous, axillary. They are unisexual, monoecious flowers, male flowers having 1-3 sessile stamens, and female flowers are solitary in nature. **Fruits:** very small, depressed globose, and moreover capsule is smooth, 2-3 mm in diameter. **Root:** branches and large [19].

### Taxonomical classification

The taxonomy of *P. niruri* is in the Kingdom (Plantae); Subkingdom (Viridiplantae); Infrakingdom (Streptophyta); Superdivision (Embryophyta); Division (Tracheophyta); Subdivision (Spermatophytina); Class (Magnoliopsida); Superorder (Rosanae); Order (Malpighiales); Family (Phyllanthaceae); Genus (*Phyllanthus*); Species (*P. niruri*) [20]. The plant genus *Phyllanthus* is a large genus of the family Phyllanthaceae,

which comprises over 800 species distributed in tropical and subtropical regions [21]. The genus name “*Phyllanthus*” means “leaf and flower” because the flower, as well as the fruit, seem to become one with the leaf [22].

### Nomenclature

*P. niruri* is a medicinal herb found mainly in rainforests of tropical areas throughout the world, including India, China, and Southeast Asia [23]. The vernacular name of *P. niruri* is also known as seed under leaf or stone breaker (English), noar (Bemgali), quebra pedra, arranca pedra (Brazil), ye xia zhu, be bei cai, xiao fan hun (Chinese), quinine creole (French), bhuiaonla, chalmeri, harfarauri (Hindi), arinelli, kizhanelli, nellipuli (Malayalam), rayavali, bhuiavli (Marathi), quebra pedra (Portuguese), ajata, amala, sukshmadala, vituntika (Sanskrit), chanca piedra, quinine criolla (Spanish), sampa-sampalukan (tagalog), keezha nelli (Tamil), nela usiri, ratsavusirike (Telugu), de sampablo (Thai) and bhui amla (Vietnam) [19,24].

### Phytochemical substances

*P. niruri* contained active phytochemical substances as follows: (1) Flavonoids including geranin, rutin, quercetin, quercitrin, astragalin, and catechin [25-29]; (2) Terpenes and terpenoids including limonene, cymene, phyllanthanol, and lupeol; (3) Coumarins including ellagic acid and methyl brevifolincarboxylate; (4) Tannins including repandusinic acid, geraniin, hypophyllanthin,

phyllanthin, phylltetralin and corilagin [30-32]; (5) Lignans [33,34]; (6) Saponins [35]; (7) Alkaloids [36].

### Traditional uses

From review literatures that regarding the traditional uses or phytochemical properties of *P. niruri* are shown in Table 1.

Phytochemical properties
Hair growth promoting activity [37]
Anti-allergic activity [32]
Anticancer activity [38-40]
Antidiabetic activity [41,42]
Anti-hepatitis B [43-45]
Anti-inflammation [46]
Anti-lipidemia activity [47,48]
Antimicrobial activity [49-52]
Antioxidant activity [53-56]
Anti-parasitic activity [57,58]
Antihyperuricemic activity [59,60]
Cardioprotective activity [61]
Hepatoprotective activity [62-68]
Nephrolithiasis treatment [69,70]
Neuroprotective activity [71]
Urolithiasis treatment [72]

### Stonebreaker activity

The Spanish name of this plant, *chanca piedra*, means "stone breaker" or "shatter stone" because it effectively uses the generation of people to eliminate gallstones and kidney stones. In Brazil, this plant is known as *quebra pedra* or *arranca pedra* which also mean to "break stone" [73]. The present review is to introduce *P. niruri* as a medicinal plant by highlighting its traditional applications in stonebreaker activity.

### In vitro study

Barros et al. (2003) [74] evaluated the effect of an aqueous extract *P. niruri* on calcium oxalate crystallization in vitro. Calcium oxalate precipitation was induced by the addition of 0.1 M sodium oxalate to unfiltered urine sample from Wistar rats and human urine in the presence or absence of *P. niruri* extract in a dose 0.25 mg/ml of urine. They observed that the presence of *P. niruri* extract did not inhibit calcium oxalate precipitation, but the crystals were significantly smaller than those in the urine samples without *P. niruri*. Moreover, they observed that after 24 hours, the

precipitated crystals formed large agglomerates in untreated urine, but the crystals remained dispersed in urine with *P. niruri*. Those authors concluded that *P. niruri* did not decrease the number of crystals but induced a marked reduction of particle size and crystal aggregation. Khare et al. (2014) [17] reported the aqueous extract of *P. niruri* could dissolve 56.8% calcium oxalate crystals when compared with cystone, the standard drug, which could dissolve 58.4% crystals in vitro study. Moreover, the aqueous extract of this plant could also inhibit up to 53.09% aggregation of calcium oxalate crystals compared to the cystone with 76.54%. Similarly, Agarwal and Varma (2014) [75] reported that the maximum inhibition was 61.97% of the aqueous extract of *P. niruri* on calcium oxalate crystallization in vitro.

### In vivo study

Freitas et al. (2002) [76] studied the effect of 42 days of chronic oral administration of 1.25 mg/ml/day *P. niruri* in adult male Wistar rats that induced urolithiasis by the introduction of a calcium oxalate crystal into the bladder. They reported that treatment with *P. niruri* strongly inhibited the growth of the matrix calculus and reduced the number of stone satellites (0.028 g) compared with the control group (0.174 g). Barros et al. (2006) [72] evaluated the effect of 30 days chronic oral administration of 5 mg/rat/ day of *P. niruri* on the performed calculus induced by introduction of a calcium oxalate seed into the bladder of male Wistar rats. They reported that *P. niruri* reduced the number and the weight of calculi compared with the untreated group. Calculi from the *P. niruri* treated group had a smoother, homogeneous surface than the spicule shape of calculi found in the untreated group. Singh et al. (2014) [77] evaluated the effect of 30 days of chronic oral administration of 150 mg/kg of *P. niruri* in ethylene glycol-induced urolithiasis in Wistar rats. Oxalate and calcium excretions significantly increased in hyperoxaluric rats as compared to control rats. The *P. niruri* treatment also significantly reduced the level of malondialdehyde and improved the activity of antioxidant enzymes, followed by a reduction of blood urea nitrogen, urea and serum creatinine. Histological analysis indicated that the *P. niruri* treatment also inhibited calcium oxalate crystal deposition and renal cell damage.

### Clinical study

Nishiura et al. (2004) [78] studied the effects of *P. niruri* on the chemical promoters and inhibitors of stone formation in known calcium stone-forming patients.

Sixty-nine patients (38±8 years old) were randomized to take either 450 mg capsules three times daily of *P. niruri* (n=33) or placebo (n=36) for 3 months. They found that there was no significant difference in urinary levels of measured metabolites calcium, uric acid, citrate, oxalate, and magnesium among all patients. However, *P. niruri* significantly reduced the mean urinary calcium in hypercalciuric patients. Micali et al. (2006) [79] studied the effect of *P. niruri* on 150 patients who had extracorporeal shock wave lithotripsy. Seventy-eight patients received *P. niruri* for 3 months or more after this lithotripsy, while 72 received none. Those with lower pole stones who received this plant had fewer recurrent lithogenesis over 6 months than those who did not use this plant. Stones in mid or upper-pole did not show significant recurrence rate changes compared with the control group.

### Phytochemical study

Grases et al. (2009) [80] evaluated the antilithiasic activity of a traditional Mallorcan herbal extract and antioxidant flavonoids, catechin, and epicatechin in Wistar rats with ethylene glycol induced calcium oxalate lithiasis. These flavonoids showed the ability to prevent the development of papillary and intratubular calcification in the kidney. Among the phytochemical substances in *P. niruri*, triterpenes have been found to inhibit the per-oxidative changes and cytotoxicity induced by calcium oxalate [81] as well as to reduce excretion of stone-forming constituents such as calcium, oxalate, and uric acid [82].

### Antilithiasic study in other *Phyllanthus* spp.

Woottisin et al. (2011) [83] evaluated the effect of 28 days chronic oral administration of 3.5 mg daily of *P. amarus* in glycolate diet induced hyperoxaluria in Wistar rats. The *P. amarus* group revealed less calcium oxalate crystal deposition in the kidney. Moreover, the stone breaker was also reported in *P. tenellus* [84]; *P. emblica* [85].

## CONCLUSION

In conclusion, even in the gallbladder, kidney, or urinary system, stone formation is a complex process that results from several events, including supersaturation, stone growth, aggregation, and retention within the tissue or organ [8]. This plant has proved to have the capacity to interfere with many stages of stone formation, reducing crystals aggregation [74], modifying their structure, shape, and composition [72,76], and altering the interaction of the crystals with

tubular cells leading to reduced subsequent endocytosis [70], as well as showed great potential to dissolve the existing stone crystal [17]. The current reviews summarized here suggest that *P. niruri* may have therapeutic potential for stonebreakers since it is able to modify the shape and texture of calculi to a smoother and probably more fragile form, which could contribute to the elimination and dissolution of calculi.

## REFERENCES

1. Romero V, Akpınar H, Assimos DG. (2010). Kidney stones: A global picture of prevalence, incidence, and associated risk factors. *Rev Urol.* 2010;12:e86-e96.
2. Scales CD, Smith AC, Hanley JM, Saigal CS. Prevalence of kidney stones in the United States. *Eur Urol.* 2012;62:160-165.
3. Richman K, O'Bell J, Pareek G. The growing prevalence of kidney stones and opportunities for prevention. *Rhode Island Med J.* 2014;12:31-34.
4. Saigal CS, Joyce G, Timilsina AR. Direct and indirect costs of nephrolithiasis in an employed population: opportunity for disease management? *Kidney Int.* 2005;68:1808-1814.
5. Antonelli J, Maalouf N, Pearle M, Lotan Y. Use of the National Health and Nutrition Examination Survey to calculate the impact of obesity and diabetes on cost and prevalence of urolithiasis in 2030. *Eur Assoc Urol.* 2014;66:724-729.
6. Preminger GM, Tiselius HG, Assimos DG, Alken P, Buck C, Gallucci M, Knoll T, Lingeman JE, Nakada SY, Pearle MS, Sarica K, Turk C, Wolf J. 2007 Guideline for the management of ureteral calculi. *J Urol.* 2007;178:2418-2434.
7. Lingeman JE, McAteer JA, Gnessin E, Evan AP. Shock wave lithotripsy: advances in technology and technique. *Nat Rev Urol.* 2009;6:660-670.
8. Ankur C, Amarchand P, Aadarsh C, Deepa I, Pawar R, Patil U. Potential of medicinal plants in kidney, gall and urinary stones. *Int J Drug Develop Res.* 2010;2:431-447.
9. Zahid IH, Bawazir AS, Naser R. Plant based native therapy for the treatment of kidney stones in Aurangabad (M.S). *J Pharmacog Phytochem.* 2013;1:189-193.
10. Velraj M, Ravichandran V. Evaluation of diuretic and anti urolithiatic activity of the whole plant *Abutilon indicum* ethanolic extract in albino rats. *Int J Pharm Sci Rev Res.* 2015;32:227-230.
11. Khan MA, Pradhan D. Antiuro lithic activity of *Ageratum conyzoides* extract in rats. *Pharmacologyonline.* 2011;3:953-958.
12. Chitra V, Gowri K, Udayasri N. Antilithiatic activity of alcoholic extract of *Boerhaavia diffusa* roots on ethylene glycol induced lithiasis in rats. *Int J Pharm Pharm Sci.* 2012;4:149-153.
13. Godara D, Kaushik V, Sharma G, Saini V. A novel herbal composition of aqueous extract of *Capparis decidua* and

- its antinephrolithiasis activity. *Am J Adv Drug Deliver.*2014;2:767-775.
14. Patil AP, Kanase SS, Naikwade NS, Disouza JL, Burli SC. Pharmacological screening of ethanolic extract of plant "Cassia tora" Linn. leaves on nephrolithiasis rats. *Int J Pharmamedix India* 2013;1:529-538.
  15. Makasana A, Ranpariya V, Desai D, Mendpara J, Parekh V. Evaluation for the anti-urolithiatic activity of *Launaea procumbens* against ethylene glycol-induced renal calculi in rats. *Toxicol Reports* 2014;1:46-52.
  16. Maya S, Pramod C. Evaluation of anti-nephrolithiatic activity of ethanolic leaf extract of *Morus alba* L. in animal models. *Int Res J Pharm.*2014;5:427-433.
  17. Khare P, Mishra VK, Arun K, Bais N, Singh R. Study on in vitro anti-lithiatic activity of *Phyllanthus niruri* Linn. leaves by homogenous precipitation and turbiditory method. *Int J Pharm Pharm Sci.* 2014;6:124-127.
  18. Patel P, Patel M, Saralai M, Gandhi T. Antiurolithiatic effects of *Solanum xanthocarpum* fruit extract on ethylene glycol induced nephrolithiasis in rats. *J Young Pharm.* 2012;4:164-170.
  19. Narendra K, Swathi J, Sowjanya KM, Krishna SA. *Phyllanthus niruri*: A review on its ethno botanical, phytochemical and pharmacological profile. *J Pharm Res.*2012;5:4681-4691.
  20. Integrated Taxonomic Information System. *Phyllanthus niruri*. Taxonomic Serial No.:504353. Geological Survey, VA, USA. 2016.
  21. Manissorn J, Sukrong S, Ruangrunsi N, Mizukami H. Molecular phylogenetic analysis of *Phyllanthus* species in Thailand and the application of polymerase chain reaction-restriction fragment length polymorphism for *Phyllanthus amarus* identification. *Biol Pharm Bull.* 2010;33:1723-1727.
  22. Uka CJ, Okeke CU, Awomukwu DA, Aziagba B, Muoka R. Taxonomic significance of foliar epidermis of some *Phyllanthus*. *IOSR J Pharm Biol Sci.* 2014;9:1-6.
  23. Bagalkotkar G, Sagineedu SR, Saad MS, Stanslas J. Phytochemicals from *Phyllanthus niruri* Linn. and their pharmacological properties: a review. *J Pharm Pharmacol.* 2006;58:1559-1570.
  24. Blake S. *Medicinal Plant Names*. LifeLong Press. 2004.
  25. Chauhan JS, Sultan M, Srivastava SK. Two new glycoflavones from the roots of *Phyllanthus niruri*. *Planta Med.* 1997;32:217-222.
  26. Gupta D, Ahmed B, Shoyakugaku Z. A new flavone glycoside from *Phyllanthus niruri*. *J Nat Prod.* 1984;383:231-215.
  27. Kotani M, Matsumoto M, Fujita A, Higa S, Wang W, Suemura M, Kishimoto T., Tanaka T. Persimmon leaf extract and astragalin inhibit development of dermatitis and IgE elevation in NC/Nga mice. *J Allergy Clin Immunol.* 2000;106:159-166.
  28. Ahmeda A, Amzad Hossain M, Ismail Z. Antioxidant properties of the isolated flavonoids from the medicinal plant *Phyllanthus niruri*. *Asian J Food Agro-Industry.* 2009;2:373-381.
  29. Jose J, Sudhakaran S, Sumesh Kumar TM, Jayaraman S, Jayadevi Variyar E. Study of in vitro immunomodulatory effect of flavonoid isolated from *Phyllanthus niruri* on human blood lymphocytes and evaluation of its antioxidant potential. *Int J Pharmacog Phytochem Res.*2014;6:284-289.
  30. Markorn M, Hasan M, Wan Daud WR, Singh H, Jahim J. Extraction of hydrolysable tannins from *Phyllanthus niruri* Linn.: Effects of solvents and extraction methods. *Sep Purif Technol.* 2007;52:487-496.
  31. Markorn M, Hasan M, Wan Daud WR, Anuar N, Hassan O, Singh H. Chemical profiling and quantification of tannins in *Phyllanthus niruri* Linn. fractionated by SFE method. *Sep Sci Technol.* 2010;46:71-78.
  32. Moreira J, Klein-Junior LC, Cechinel Filho V, Campos Buzzi F. Anti-hyperalgesic activity of corilagin, a tannin isolated from *Phyllanthus niruri* L. (Euphorbiaceae). *J Ethnopharmacol.*2013;146:318-323.
  33. Anjaneyulu A, Rao J, Row L, Subrahmanyam C. Crystalline constituents of Euphorbiaceae-XII: Isolation and structural elucidation of three new lignans from the leaves of *Phyllanthus niruri*. *Tetrahedron.* 1973;29:1291-1298.
  34. Elfahmi Y, Batterman S, Koulman A, Hack T, Bos R, Kayser O, Woerdenbag H, Quax W. Lignans from cell suspension cultures of *Phyllanthus niruri*, an Indonesian medicinal plant. *J Nat Prod.* 2006;69:55-58.
  35. Ajibade V, Famurewa O. Histopathological and toxicological effects of crude saponin extract from *Phyllanthus niruri*, L (Syn. *P. franternus*. Webster) on organs in animal studies. *Global J Med Res.* 2012;12:31-37.
  36. Mulchandani NB, Hasarajani SA. 4-methoxy-nor-securinine, a new alkaloid from *Phyllanthus niruri*. *Planta Med.* 1984;50:104-105.
  37. Patel S, Sharma V, Chauhan S, Thakur M, Dixit VK. Evaluation of hair growth promoting activity of *Phyllanthus niruri*. *Avicenna J Phytomed.* 2015;5:512-519.
  38. Sharma P, Parmar J, Verma P, Sharma P, Goyal P. Anti-tumor activity of *Phyllanthus niruri* (a medicinal plant) on chemical-induced skin carcinogenesis in mice. *Asian Pac. J. Cancer Prev.* 2009;10:1089-1094.
  39. Sharma P, Parmar J, Verma P, Goyal PK. Modulatory influence of *Phyllanthus niruri* on oxidative stress, antioxidant defense and chemically induced skin tumors. *J Environ Pathol Toxicol Oncol.* 2011;30:43-53.
  40. Araujo Junior RF, Souza TP, Pires JG, Soares LA, Araujo AA, Petrovick PR, Macedo HD, Leitao Oliveira AL, Guerra GC. A dry extract of *Phyllanthus niruri* protects normal cells and induces apoptosis in human liver carcinoma cells. *Exp Biol Med.* 2012;237:1281-1288.
  41. Okoli CO, Obidike IC, Ezike AC, Akah PA, Salawu OA. Studies on the possible mechanisms of antidiabetic activity of extract of aerial parts of *Phyllanthus niruri*. *Pharm Biol.* 2011;49:248-255.
  42. Giribabu N, Rao PV, Kumar KP, Muniandy S, Swapna Rekha S, Salleh N. Aqueous extract of *Phyllanthus niruri* leaves displays in vitro antioxidant activity and prevents the elevation of oxidative stress in the kidney of streptozotocin-induced diabetic male rats. *Evid Based Complement Alternat Med.*2014;2014:834815.



43. Wei W, Li X, Wang K, Zheng Z, Zhou M. Lignans with anti-hepatitis B virus activities from *Phyllanthus niruri* L. *Phytother Res*. 2012;26:964-968.
44. Liu S, Wei W, Li Y, Lin X, Shi K, Cao X, Zhou M. In vitro and in vivo anti-hepatitis B virus activities of the lignan nirtetralin B isolated from *Phyllanthus niruri* L. *J Ethnopharmacol*. 2014;157:62-68.
45. Mohan M, James P, Vasalan R, Nazeem PA. Molecular docking studies of phytochemicals from *Phyllanthus niruri* against hepatitis B DNA polymerase. *Bioinformation* 2015;11:426-431.
46. Melo MN, Soares LA, Porto CR, Araujo AA, Almeida M, Souza TP, Petrovick PR, Araujo RF, Guerra GC. Spray-dried extract of *Phyllanthus niruri* L. reduces mucosal damage in rats with intestinal inflammation. *J Pharm Pharmacol*. 2015;67:1107-1118.
47. Khanna AK, Rizvi F, Chander R. Lipid lowering activity of *Phyllanthus niruri* in hyperlipemic rats. *J Ethnopharmacol*. 2002;82:19-22.
48. Latha P, Chaitanya D, Rukkumani R. Protective effect of *Phyllanthus niruri* on alcohol and heated sunflower oil induced hyperlipidemia in Wistar rats. *Toxicol Mech Methods* 2010;20:498-503.
49. Mustofa J, Shoikhah E, Wahyuono S. In vitro and in vivo antiparasitic activity and cytotoxicity of extracts of *Phyllanthus niruri* L. herbs traditionally used to treat malaria in Indonesia. *Southeast Asian J Trop Med Pub Health*. 2007;38:609-615.
50. Amin Z, Abdulla M, Ali H, Alshawsh M, Qadir S. Assessment of in vitro antioxidant, antibacterial and immune activation potentials of aqueous and ethanol extracts of *Phyllanthus niruri*. *J Sci Food Agric*. 2012;92:1874-1877.
51. Ranilla LG, Apostolidis E, Shetty K. Antimicrobial activity of an Amazon medicinal plant (*Chancapiedra*) (*Phyllanthus niruri* L.) against *Helicobacter pylori* and lactic acid bacteria. *Phytother. Res*. 2012;26:791-799.
52. Ibrahim D, Hong L, Kuppan N. Antimicrobial activity of crude methanolic extract from *Phyllanthus niruri*. *Nat Prod Commun*. 2013;8:493-496.
53. Harish R, Shivanandappa T. Antioxidant activity and hepatoprotective potential of *Phyllanthus niruri*. *Food Chem*. 2006;95:180-185.
54. Sarkar MK, Sil PC. Prevention of tertiary butyl hydroperoxide induced oxidative impairment and cell death by a novel antioxidant protein molecule isolated from the herb, *Phyllanthus niruri*. *Toxicol. In Vitro* 2010;24:1711-1719.
55. Bhattacharyya S, Ghosh S, Sil PC. Amelioration of aspirin induced oxidative impairment and apoptotic cell death by a novel antioxidant protein molecule isolated from the herb *Phyllanthus niruri*. *PLoS One*. 2014;9:e89026.
56. Colpo E, Vilanova CD, Pereira RP, Reetz LG, Oliveira L, Farias IL, Boligon AA, Athayde ML, Rocha JB. Antioxidant effects of *Phyllanthus niruri* tea on healthy subjects. *Asian Pac J Trop Med*. 2014;7:113-118.
57. Shakil NA, Pankaj Kumar J, Pandey RK, Saxena DB. Nematicidal prenylated flavanones from *Phyllanthus niruri*. *Phytochemistry* 2008;69:759-764.
58. Ifeoma O, Samuel O, Itohan AM, Adeola SO. Isolation, fractionation and evaluation of the antiparasitic properties of *Phyllanthus niruri* resident in its chloroform fraction. *Asian Pac J Trop Med*. 2013;6:169-175.
59. Murugaiyah V, Chan KL. Antihyperuricemic lignans from the leaves of *Phyllanthus niruri*. *Planta Med*. 2006;72:1262-1267.
60. Murugaiyah V, Chan KL. Mechanisms of antihyperuricemic effect of *Phyllanthus niruri* and its lignan constituents. *J Ethnopharmacol*. 2009;124:233-239.
61. Thippeswamy AH, Shirodkar A, Koti BC, Sadiq AJ, Praveen DM, Swamy AH, Patil M. Protective role of *Phyllanthus niruri* extract in doxorubicin-induced myocardial toxicity in rats. *Indian J Pharmacol*. 2011;43:31-35.
62. Chatterjee M, Sarkar K, Sil PC. Herbal (*Phyllanthus niruri*) protein isolate protects liver from nimesulide induced oxidative stress. *Pathophysiol*. 2006;13:95-102.
63. Bhattacharjee R, Sil PC. Protein isolate from the herb, *Phyllanthus niruri* L. (Euphorbiaceae), plays hepatoprotective role against carbon tetrachloride induced liver damage via its antioxidant properties. *Food Chem Toxicol*. 2007;45:817-826.
64. Chatterjee M, Sil PC. Protective role of *Phyllanthus niruri* against nimesulide induced hepatic damage. *Indian J Clin Biochem*. 2007;22:109-116.
65. Sabir SM, Rocha JBT. Water-extractable phytochemicals from *Phyllanthus niruri* exhibit distinct in vitro antioxidant and in vivo hepatoprotective activity against paracetamol-induced liver damage in mice. *Food Chem*. 2008;111:845-851.
66. Manjrekar AP, Jisha V, Baq PP, Adhikary B, Pai MM, Heqde A, Nandini M. Effect of *Phyllanthus niruri* Linn. treatment on liver, kidney and testes in CCl<sub>4</sub> induced hepatotoxic rats. *Indian J Exp Biol*. 2008;46:514-520.
67. Amin Z, Bilgen M, Alshawsh M, Ali H, Hadi A, Abdulla M. Protective role of *Phyllanthus niruri* extract against thioacetamide-induced liver cirrhosis in rat model. *Evid Based Complement Alternat Med*. 2012;2012:241583.
68. Amin Z, Alshawsh M, Kassim M, Ali H, Abdulla M. Gene expression profiling reveals underlying molecular mechanism of hepatoprotective effect of *Phyllanthus niruri* on thioacetamide-induced hepatotoxicity in Sprague Dawley rats. *BMC Complement Alternat Med*. 2013;13:160.
69. Campos AH, Schor N. *Phyllanthus niruri* inhibits calcium oxalate endocytosis by renal tubular cells: its role in urolithiasis. *Nephron*. 1999;81:393-397.
70. Boim MA, Heilberg IP, Schor N. *Phyllanthus niruri* as a promising alternative treatment for nephrolithiasis. *Int Braz J Urol*. 2010;36:657-664.
71. Obidike IC, Salawu OA, NdukubaM., Okoli CO, Osunkwo UA. The anti-inflammatory and antinociceptive properties of the chloroform fraction from *Phyllanthus niruri* plant is mediated via the peripheral nervous system. *J Diet Suppl*. 2010;7:341-350.
72. Barros ME, Lima R, Mercuri LP, Matos JR, Schor N, Boim MA. Effect of extract of *Phyllanthus niruri* on crystal deposition in experimental urolithiasis. *Urol Res*. 2006;34:351-357.

73. Taylor L. Chanca piedra "Stone Breaker" (*Phyllanthus niruri*). In: Herbal Secrets of the Rainforest. Austin: Sage Press, Inc. 2003.
74. Barros ME, Schor N, Boim MA. Effects of an aqueous extract from *Phyllanthus niruri* on calcium oxalate crystallization in vitro. *Urol Res*. 2003;30:374-379.
75. Agarwal K, Varma R. Investigating antiurolithiatic potential of *Phyllanthus niruri* L. a member of the Family Euphorbiaceae. *Am J Phytomed Clin Ther*. 2014;2:423-431.
76. Freitas AM, Schor N, Boim MA. The effect of *Phyllanthus niruri* on urinary inhibitors of calcium oxalate crystallization and other factors associated with renal stone formation. *BJU Int*. 2002;89:829-834.
77. Singh MF, Bisht S, Sajwan R. Evaluation of antiurolithiatic activity of combination of *Phyllanthus niruri* and *Aerva lanata* in ethylene glycol induced urolithiasis in rats. *Guru Drone J Pharm Res*. 2014;2:18-29.
78. Nishiura JL, Campos AH, Boim MA, Heiberg IP, Schor N. *Phyllanthus niruri* normalizes elevated urinary calcium levels in calcium stone forming (CSF) patients. *Urol Res*. 2004;32:362-366.
79. Micali S, Sighinolfi MC, Celia A, De Stefani S, Grande M, Cicero A, Bianchi G. Can *Phyllanthus niruri* affect the efficacy of extracorporeal shock wave lithotripsy for renal stones? A randomized prospective, long-term study. *J Urol*. 2006;176:1020-1022.
80. Grases F, Prieto RM, Gomila I, Sanchis P, Costa Bauza A. Phytotherapy and renal stones: The role of antioxidants. A pilot study in Wistar rats. *Urol Res*. 2009;37:35-40.
81. Malini MM, Lenin M, Varalakshmi P. Protective effect of triterpenes on calcium oxalate crystal-induced peroxidative changes in experimental urolithiasis. *Pharmacol Res*. 2000;41:413-418.
82. Vidya L, Lenin M, Varalakshmi P. Evaluation of the effect of triterpenes on urinary risk factors of stone formation in pyridoxine deficient hyperoxaluric rats. *Phytother Res*. 2002;16:514-518.
83. Woottisin S, Hossain RZ, Yachantha C, Sriboonlue P, Ogawa Y, Saito S. Effects of *Orthosiphon grandiflorus*, *Hibiscus sabdariffa* and *Phyllanthus amarus* extracts on risk factors for urinary calcium oxalate stones in rats. *J Urol*. 2011;185:323-328.
84. Silva TCL, Filho JV, Amorim ELC, Souza IA, Albuquerque UP, Araujo EC. Acute toxicity study of stone-breaker (*Phyllanthus tenellus* Roxb.) *Rev Ciênc Farm Appl*. 2012;33:205-210.
85. Bindhu B, Swetha AS, Veluraja K. Studies on the effect of *Phyllanthus emblica* extract on the growth of urinary type struvite crystals in vitro. *Clin Phytosci*. 2015;1:3.